

Geobiology 2013 Lecture 6

Biogeochemical Tracers

Isotopics #3:

Biosynthetic fractionations and Intramolecular isotopic data, more of multi-element isotopics and the Precambrian C-Cycle

Acknowledgements: John Hayes, David DesMarais

Assigned Reading

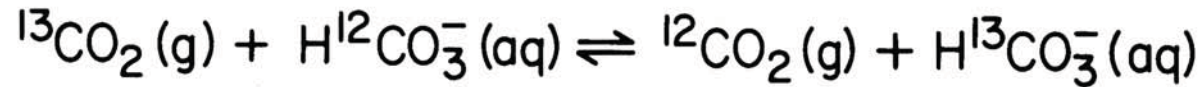
- Hayes JM 2001 Fractionation of the isotopes of carbon and hydrogen in biosynthetic processes. *Reviews in Mineralogy Stable Isotopic Geochemistry*, John W. Valley and David R. Cole (eds.)
- David J Des Marais 1997. Isotopic evolution of the biogeochemical carbon cycle during the Proterozoic Eon Original Research Article, *Organic Geochemistry*, Volume 27, Issues 5–6, Pages 185-193

Revision of carbon isotopic principles

And examining isotopic fractionation at
molecular, organismic and planetary
scales

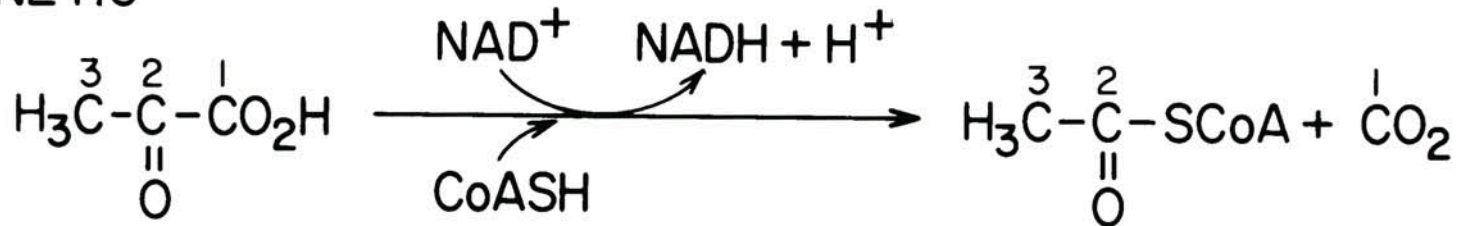
Isotope Effects

EQUILIBRIUM



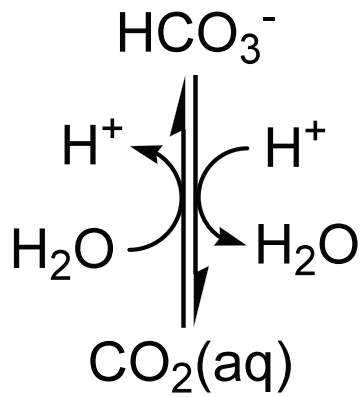
$$K = \begin{array}{ll} 1.0092 & (0^\circ\text{C}) \\ 1.0068 & (30^\circ\text{C}) \end{array}$$

KINETIC

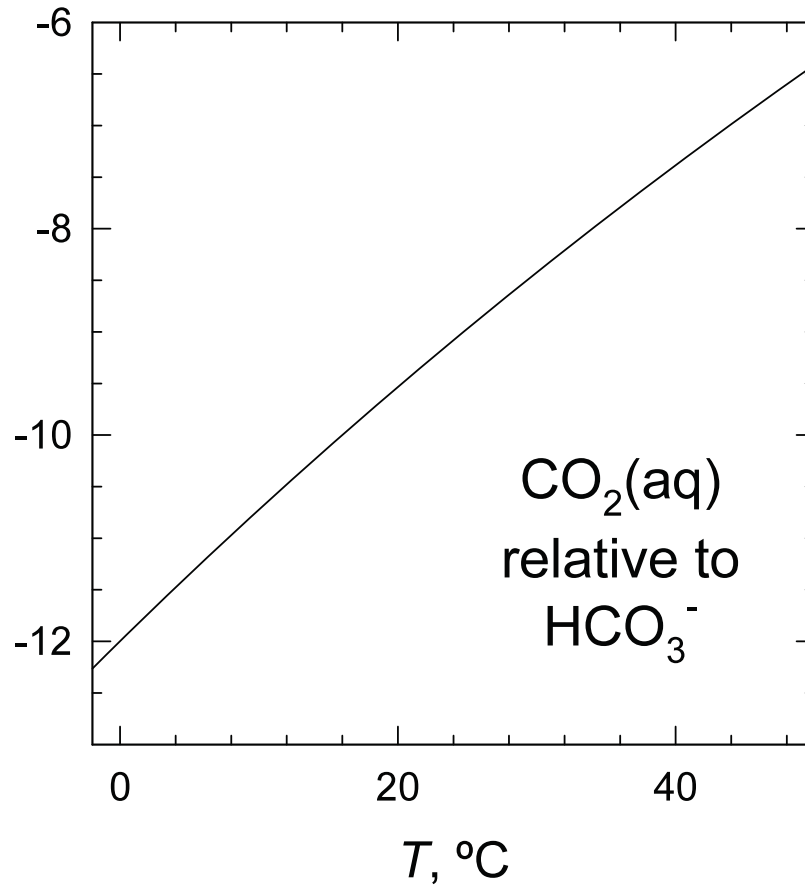


Pyruvate dehydrogenase

$$\left(\frac{{}^{12}k}{{}^{13}k} \right)_{\text{C-2}} = 1.0232$$



Mook *et al.*, 1974

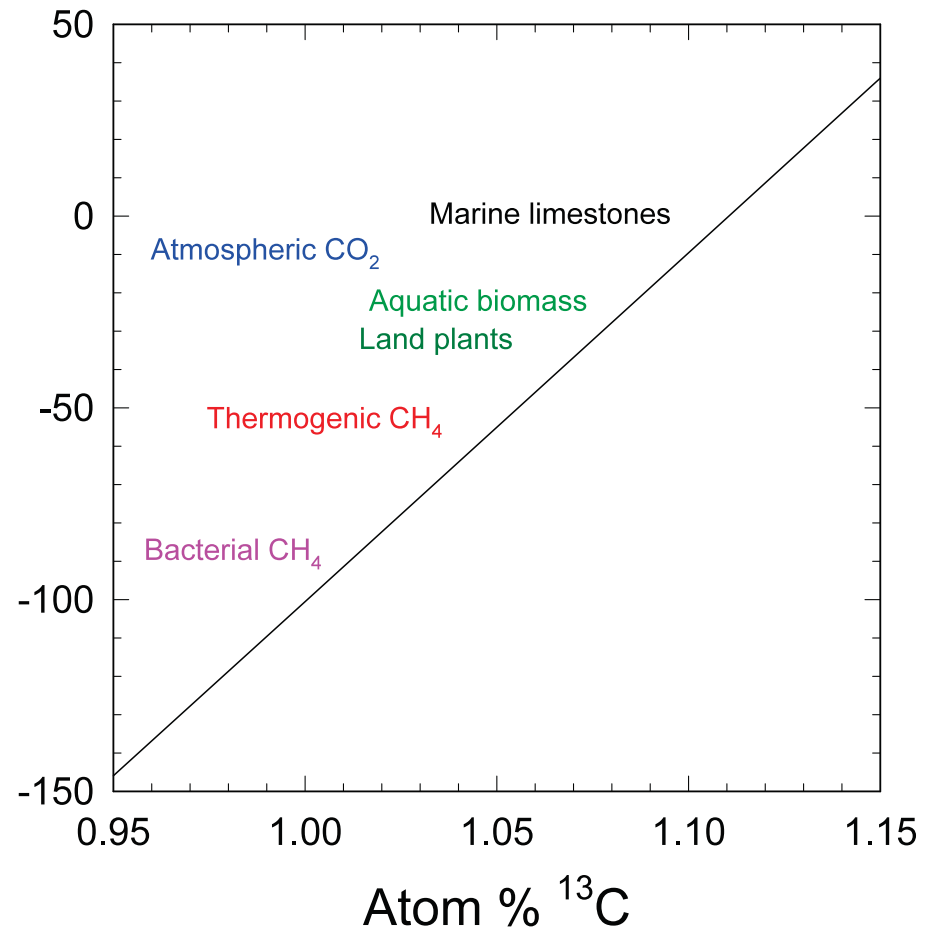


$$R \equiv \frac{^{13}\text{C}}{^{12}\text{C}} \quad \delta^{13}\text{C} = \frac{R_{\text{sample}}}{R_{\text{VPDB}}} - 1$$

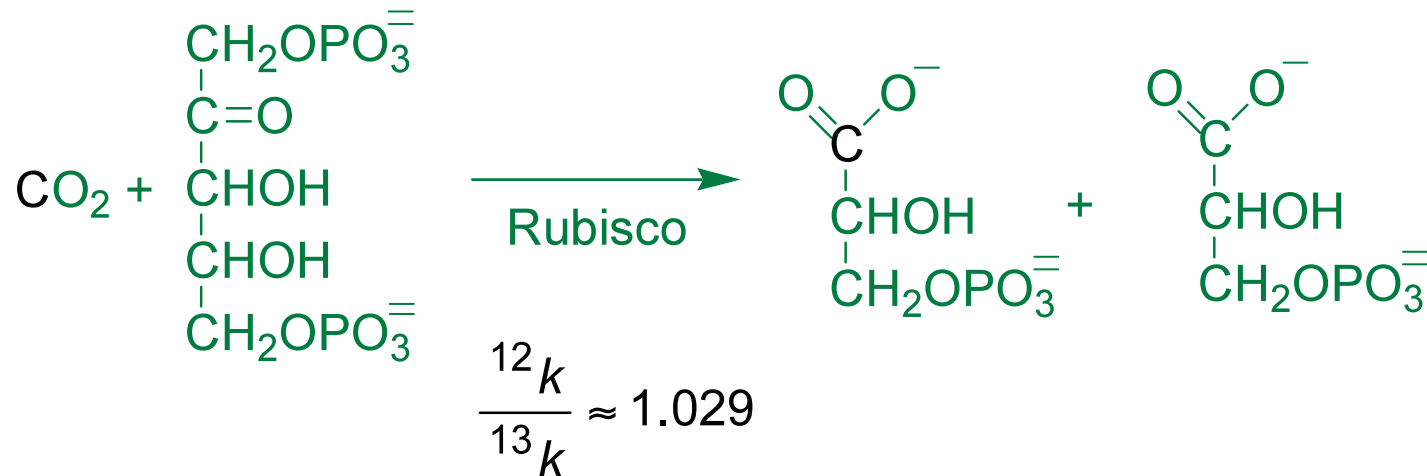
Abundances of ^{13}C are reported in terms of δ . The zero point of the scale is defined by the VPDB standard.

Values of δ are commonly multiplied by 1000 and thus expressed in parts per thousand, ‰.

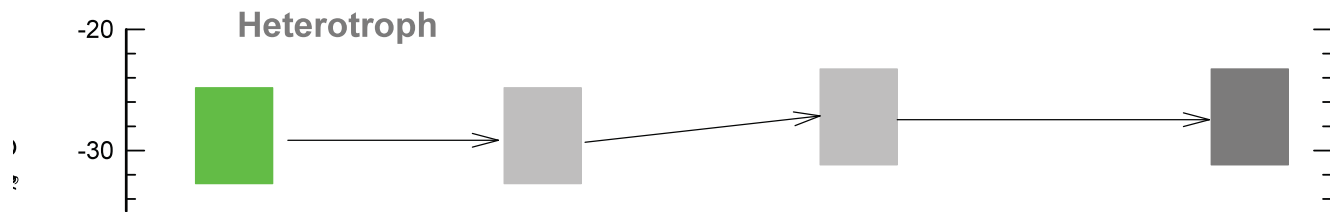
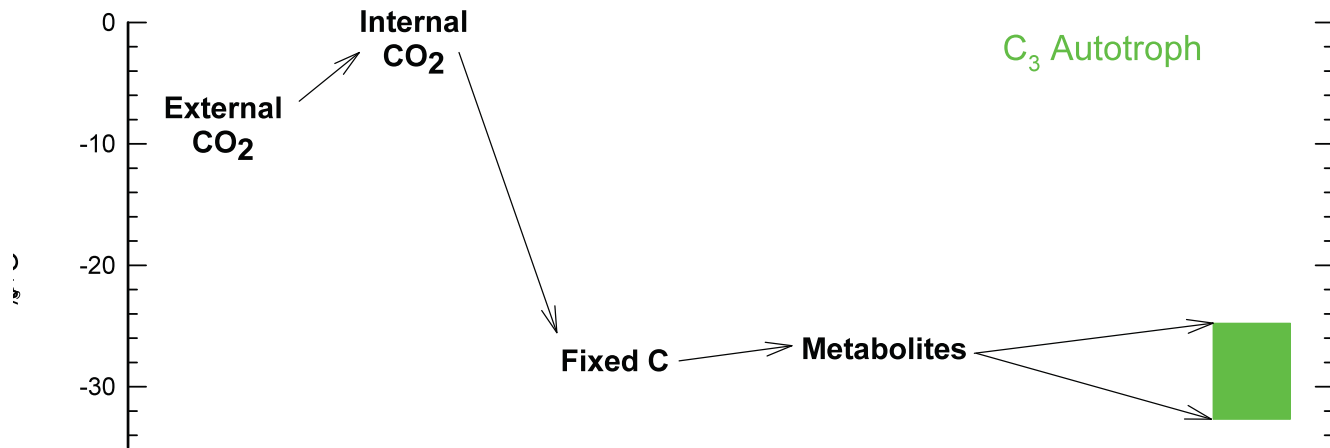
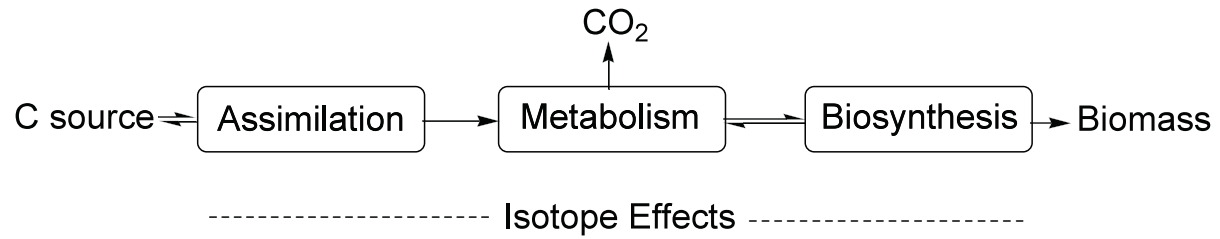
$\delta^{13}\text{C}$
‰VPDB



Isotope effects associated with biochemical reactions cause isotopic variations in nature



Fixed C will be depleted in ^{13}C relative to CO_2
 The relative depletion will be 29 parts per thousand
 e. g., if $\text{CO}_2 = 1.000\% \text{ } ^{13}\text{C}$, fixed C = $0.971\% \text{ } ^{13}\text{C}$



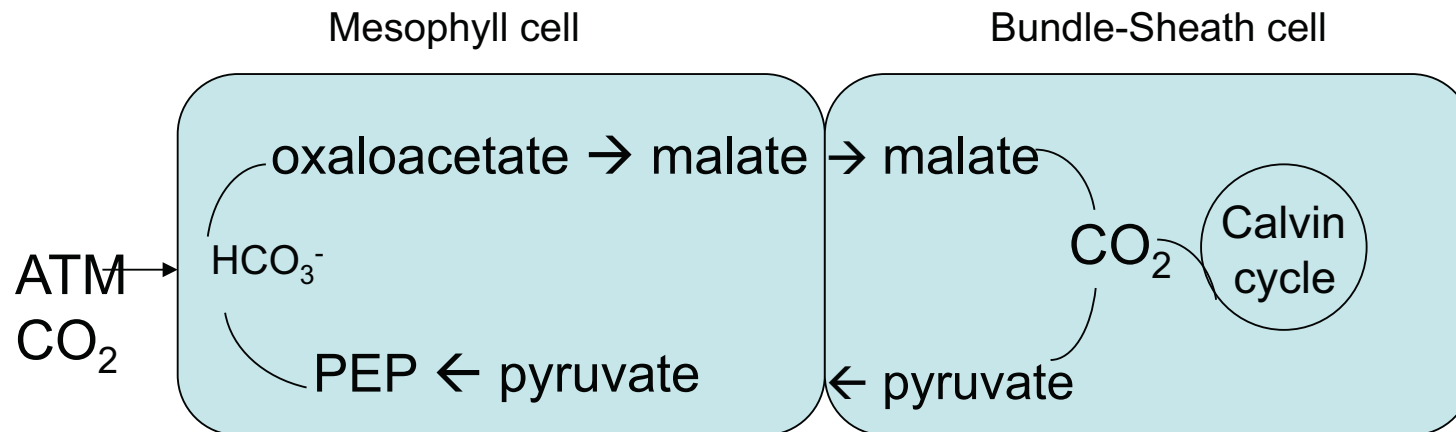
Courtesy of John Hayes. Used with permission.

Fractionation of C-Isotopes during Autotrophy

Pathway, enzyme	React & substr	Product	ϵ ‰	Organisms
C3			10-22	
Rubisco1	$\text{CO}_2 + \text{RUBP}$	3-PGA x 2	30	plants & algae
Rubisco2	$\text{CO}_2 + \text{RUBP}$	3-PGA x 2	22	cyanobacteria
PEP carboxylase	$\text{HCO}_3^- + \text{PEP}$	oxaloacetate	2	plants & algae
PEP carboxykinase	$\text{CO}_2 + \text{PEP}$	oxaloacetate		plants & algae
C4 and CAM			2-15	
PEP carboxylase	$\text{HCO}_3^- + \text{PEP}$	oxaloacetate	2	plants &
Rubisco1	$\text{CO}_2 + \text{RUBP}$	3-PGA x 2	30	algae (C4)
Acetyl-CoA			15-36	bacteria
CO dehydrog	$\text{CO}_2 + 2\text{H}^+ + \text{CoASH}$	AcSCoA	52	
Pyruvate synthase	$\text{CO}_2 + \text{Ac-CoA}$	pyruvate		
PEP carboxylase	$\text{HCO}_3^- + \text{PEP}$	oxaloacetate	2	
PEP carboxykinase	$\text{CO}_2 + \text{PEP}$	Oxaloacetate		
Reductive or reverse TCA	$\text{CO}_2 + \text{succinyl-CoA (+ others)}$	α -ketoglutarate	4-13	Bacteria esp green sulfur
3-hydroxypropionate	$\text{HCO}_3^- + \text{acetylCoA}$	Malonyl-CoA		Green non-S

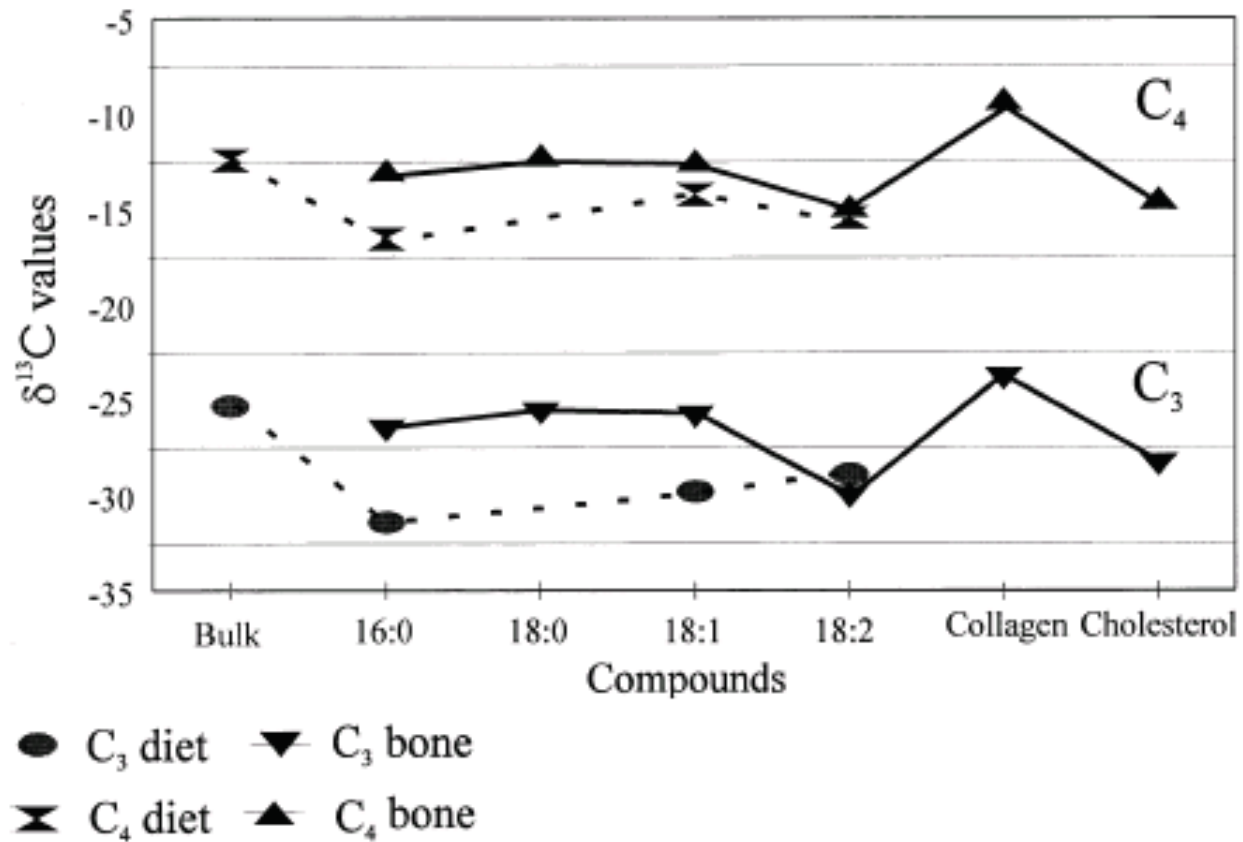
Carbon fixation (C4 & CAM pathways)

Formation of oxaloacetate from PEP (Phosphoenolpyruvate)
catalysed by PEP carboxylase



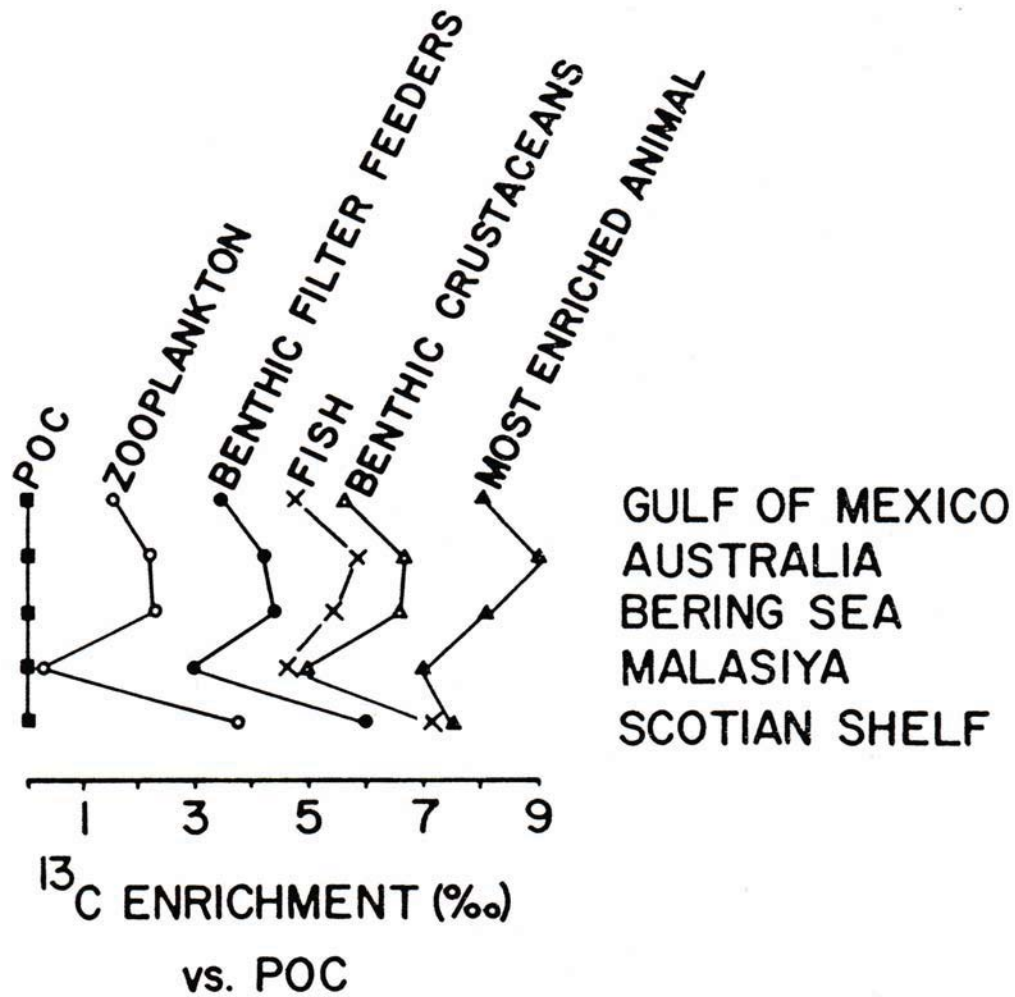
CAM (Crassulacean acid metabolism): Use both
C3 and C4 metabolism separated in time

Isotopic consequences of different food sources



A. W. Stott, E. Davies, R. P. Evershed, & N. Tuross (1997) *Naturwissenschaften* **84**, 82–86.

Trophic Shifts



Intramolecular C-isotopic Differences

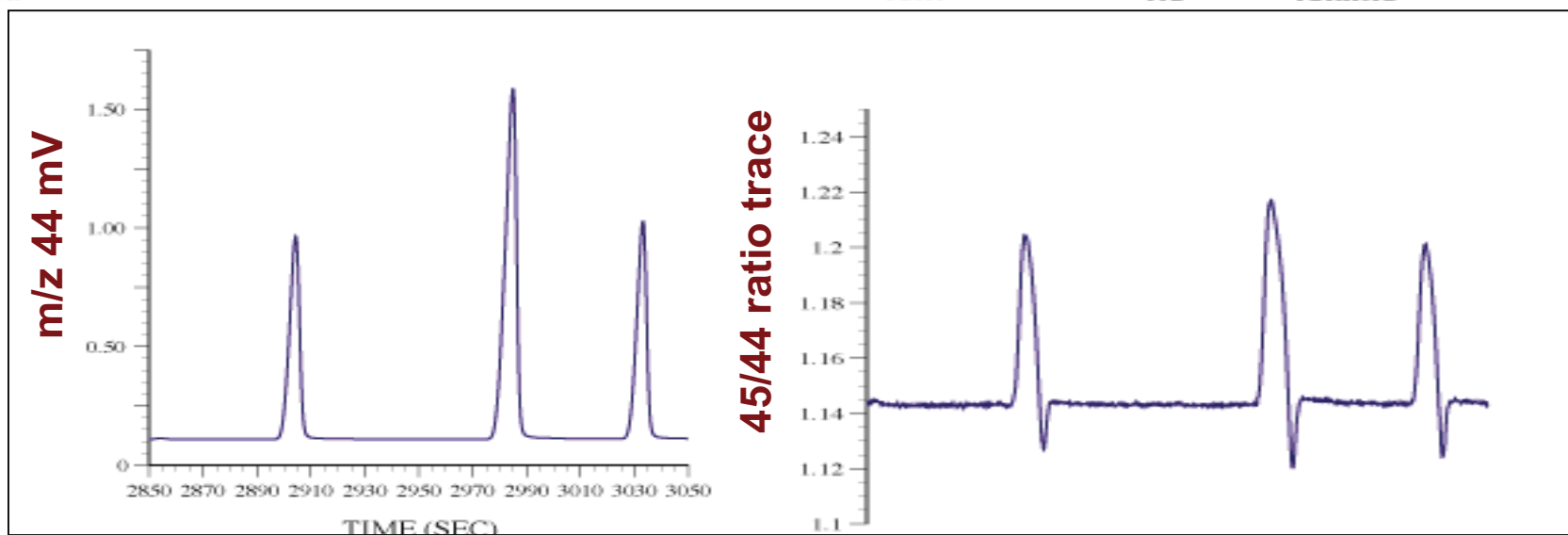
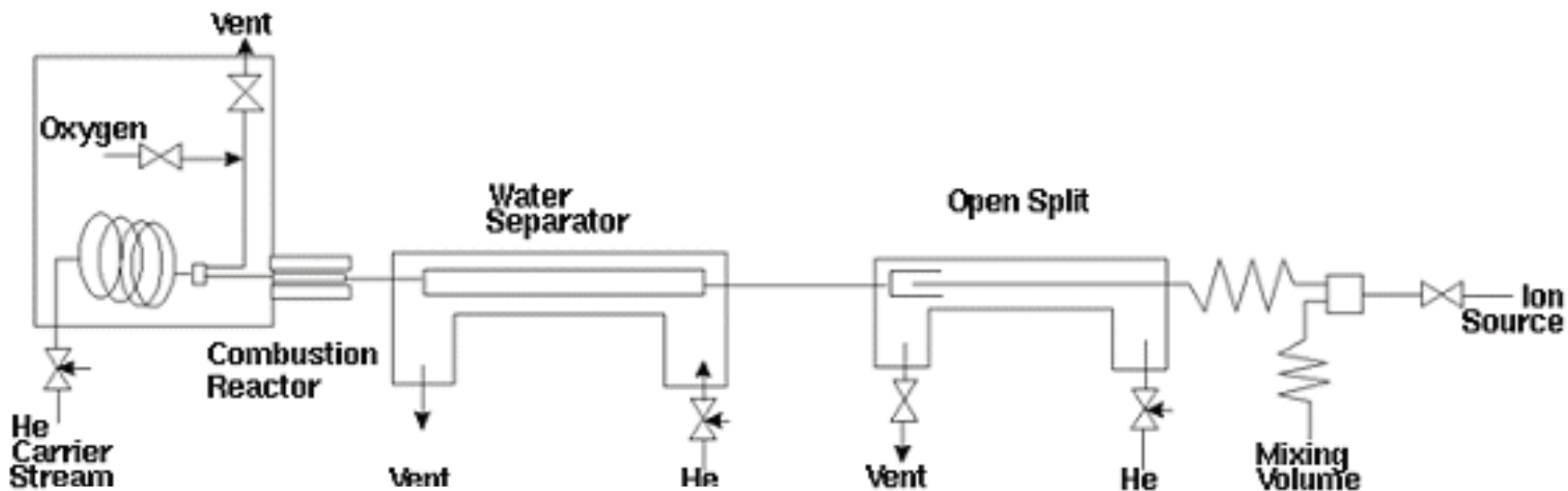
(DeNiro and Epstein, 1977; Monson and Hayes, 1980, 1982; reviewed Hayes, 2001)

Reactions occur between molecules but isotope selectivity is expressed as chemical bonds that are made or broken at particular carbon positions.

Isotope effects pertain to those specific positions and control fractionations only at that reaction site, not throughout the whole molecule.

To calculate changes in the isotopic compositions of whole molecules we must first calculate the change at the site and then allow for the rest of the molecule because the isotopic shift is diluted by mixing with carbon that is just along for the ride.....Hayes, 2002

Compound Specific Isotope Analysis



Courtesy of John Hayes. Used with permission.

TABLE 1 Carbon isotopic compositions of individual compounds

Peak	t_R^* (s)	Amount† (nmol C)	$\delta^{13}C_{\ddagger}$ (‰)	Identification
1	1,679	1.1	-22.7 ± 1.0	norpristane
2	1,722	1.0	-30.2 ± 0.3	C ₁₉ acyclic isoprenoid
3	1,812	0.7	-25.4 ± 1.0	pristane
4	2,040	2.0	-31.8 ± 0.8	phytane
5	2,602	1.0	-29.1 ± 0.6	C ₂₃ acyclic isoprenoid
6	3,161	1.3	-23.9 ± 0.6	10β(H)-des-A-lupane
7	3,571	1.3	-24.9 ± 1.0	mixture of hydrocarbons
8	3,688	2.6	-73.4 ± 1.3	C ₃₂ acyclic isoprenoid
9	3,883	0.9	-24.2 ± 1.2	isoprenoid alkane
10	3,957	6.8	-49.9 ± 1.1	17β(H)-22,29,30-trisnorhopane
11	3,977	2.0	-60.4 ± 1.8	isoprenoid alkane
12	4,100	1.6	-43.5 ± 1.0	17α(H),21β(H)-30-norhopane
13	4,156	2.0	~ -45	17β(H),21α(H)-30-norhopane§
14	4,210	2.9	~ -34	17α(H),21β(H)-hopane
15	4,256	6.2	-65.3 ± 1.4	17β(H),21β(H)-30-norhopane
16	4,364	1.8	-39.4 ± 0.8	17α(H),21β(H)-homohopane
17	4,392	1.3	-35.2 ± 1.4	17β(H),21β(H)-hopane
18	4,552	4.2	-36.6 ± 0.5	17β(H),21β(H)-homohopane
19	4,692	15.4	-20.9 ± 0.5	lycopane¶
20	5,010	0.5	-27.0 ± 0.4	unknown hydrocarbon
21	5,408	0.8	-28.8 ± 1.0	unknown hydrocarbon

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ACKNOWLEDGEMENTS. We thank A. Hildebrand for discussion of the H/T boundary in and petrology, D. Gilespie for discussion early in this study, R. Selkirk for information and G. Shoemaker for comments.

Evidence from carbon isotope measurements for diverse origins of sedimentary hydrocarbons

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Reprinted by permission from Macmillan Publishers Ltd. Katherine H. Freeman, J. M. Hayes, et al. Evidence from Carbon Isotope Measurements for Diverse Origins of Sedimentary Hydrocarbons. *Nature* 343 (1990): 254-6.

C-isotopic Composition of Individual Organic Compounds

Three major controls

- Source of carbon and its C-isotopic composition
- Fractionation during assimilation (eg heterotrophy, photosynthesis, methanotrophy)
- Fractionation during biosynthesis (lipids)

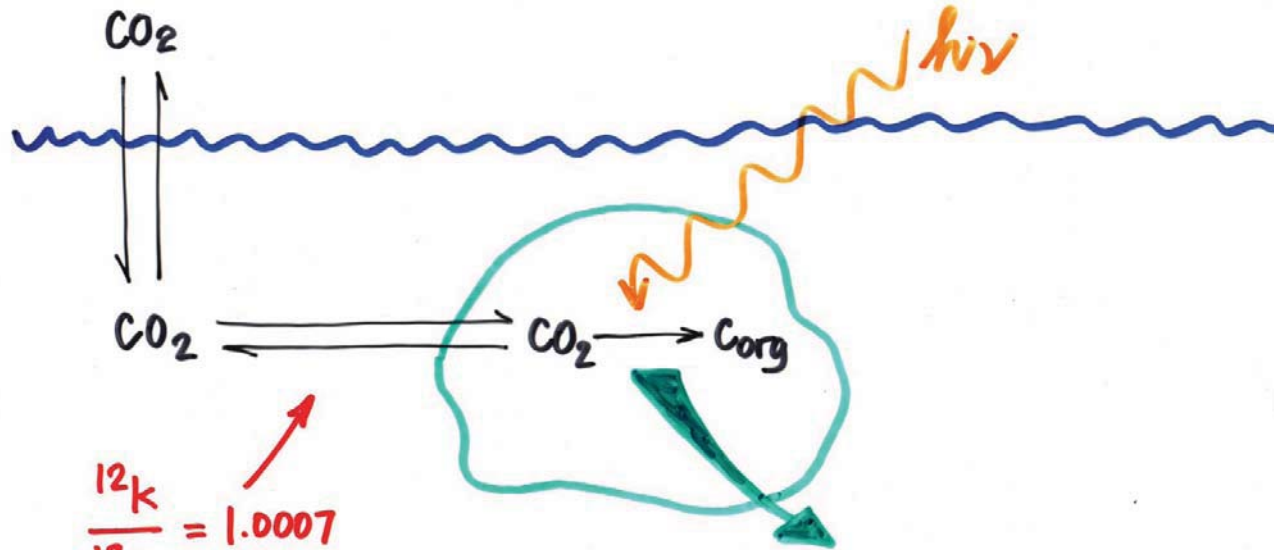
C-isotopic Composition of Organic Compounds

Source of carbon and its C-isotopic composition

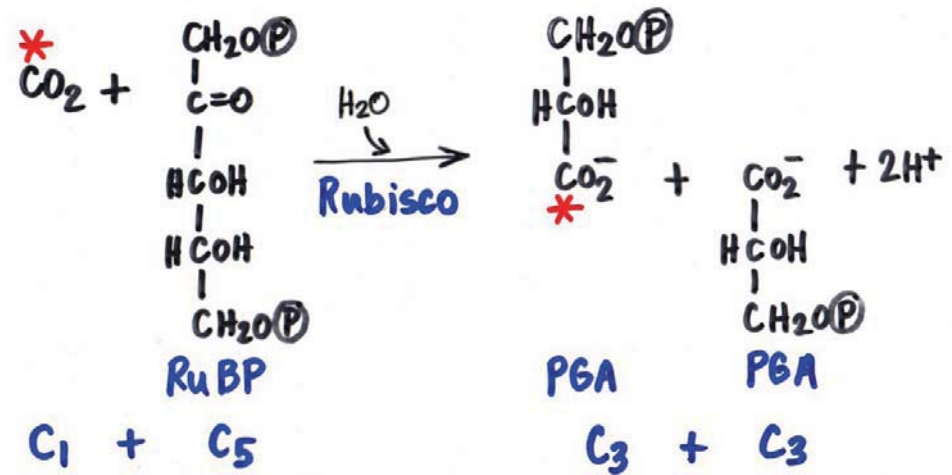
– Inorganic carbon

- (-7‰ atm. CO₂) assimilated by photosynthesis

$\epsilon \rightarrow$ 5-35 per mil depending on pathway extent of consumption



$$\frac{12\text{k}}{13\text{k}} = 1.0007$$



$$\frac{12\text{k}}{13\text{k}} = 1.029$$

C-isotopic Composition of Individual Organic Compounds

Source of carbon and its C-isotopic composition

- Inorganic carbon
 - (-7‰ atm. CO₂) assimilated by photosynthesis
 - ε → 5-35 per mil depending on pathway extent of consumption
- Organic carbon
 - (-25‰ on average) assimilated during heterotrophy
 - ε → -1 (you are what you eat plus 1 per mil!!)
- Methane carbon
 - (-30 to -100‰) assimilated during methanotrophy
 - ε → 0-30 per mil depending on pathway and extent of consumption

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Café Methane

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At the very edge of the brine pool, the mussels are especially abundant and happy. This area is often filled with newly settled baby mussels perched on the shells of larger mussels just above the brine.

Credit: Penn State University, Dept. of Biology

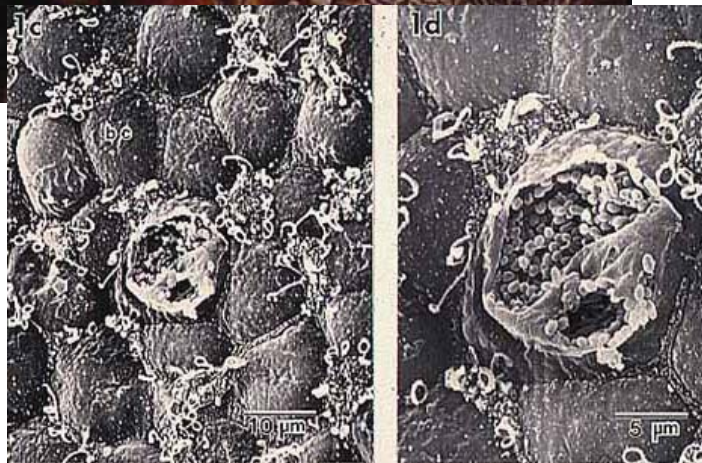
Gas hydrates (yellow) are ice with gas trapped inside; exposed beds are accessible to submersibles on the deep sea floor of the Gulf of Mexico. [Ice worms](#), a new species only seen in hydrate, were discovered in 1997 by C. Fisher, Penn State University.

Credit: I. MacDonald



Methane-rich water is pumped into the mussel and across its gills. The symbiotic bacteria in the gills use methane as both a carbon and energy source. The mussels, in turn, live off the symbiotic bacteria.

Courtesy of Charles Fisher, Penn State University, Dept. of Biology. Used with permission.



Using the scanning electron microscope, we can see over a dozen mussel gill cells in the panel on the left. On the right is a closer look at the cell with its outer membrane partially removed. Look into the cell to see hundreds of symbiotic bacteria.

Identification of Methanotrophic Lipid Biomarkers in Cold-Seep Mussel Gills: Chemical and Isotopic Analysis

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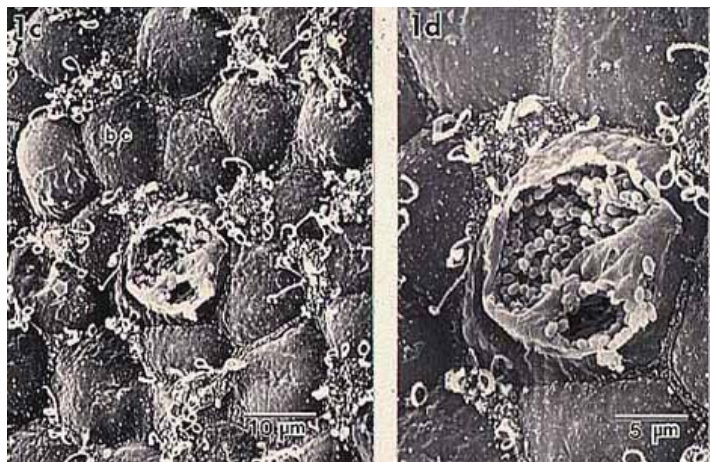


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Using the scanning electron microscope, we can see over a dozen mussel gill cells in the panel on the left. On the right is a closer look at the cell with its outer membrane partially removed. Look into the cell to see hundreds of symbiotic bacteria.

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Aerobic methanotrophy at ancient marine methane seeps: A synthesis

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ABSTRACT

The molecular fingerprints of the chemosynthesis based microbial communities at methane seeps tend to be extremely well preserved in authigenic carbonates. The key process at seeps is the anaerobic oxidation of methane (AOM), which is performed by consortia of methanotrophic archaea and sulphate reducing bacteria. Besides the occurrence of ^{13}C depleted isoprenoids and *n*-alkyl chains derived from methanotrophic archaea and sul-

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Table 1

Background information on samples

Location	Fossil inventory	Carbonate microfabrics	Stable isotopes	References
Tepee Buttes (Campanian)	Lucinid bivalves, gastropods	Clotted micrite, yellow calcite, banded/botryoidal cement, <i>in situ</i> brecciation	Yellow calcite $\delta^{13}\text{C}$: -45.9 to -31.7‰, micrite $\delta^{13}\text{C}$: -49.7 to -43.1‰, banded/botryoidal cement $\delta^{13}\text{C}$: -45.5 to -13.2‰	Kauffman et al., 1996; Shapiro, 2004; Birgel et al., 2006b
Pietralunga (Miocene)	Lucinid bivalves	Microcrystalline calcite, aragonitic cement, fossilized filaments	Microcrystalline calcite $\delta^{13}\text{C}$: -51.5 to -45.8‰, aragonitic cement $\delta^{13}\text{C}$: -43.3 to -40.8‰	Peckmann et al., 2004; Barbieri and Cavalazzi, 2005
Marmorito (Miocene)	None	Microcrystalline dolomite, calcitic veins, <i>in situ</i> brecciation	Microcrystalline dolomite $\delta^{13}\text{C}$: -40.7 to -38.9‰, calcitic vein $\delta^{13}\text{C}$: -28.5 to -17.3‰	Clari et al., 1988; Peckmann et al., 1999

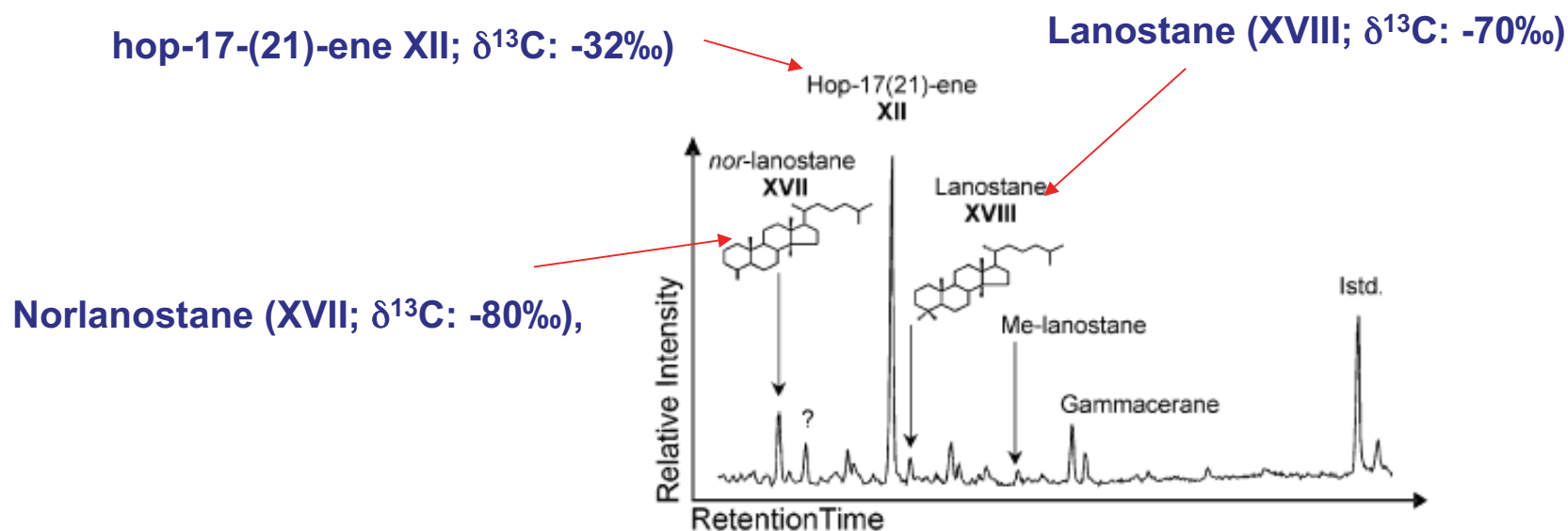
 $\delta^{13}\text{C}$ carbonate values in ‰ relative to V-PDB

Fig. 1. Partial gas chromatogram (total Ion Current: TIC) of hydrocarbon fraction from Pietralunga. Istd.: internal standard. Roman numbers: see Appendix.

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C-isotopic Composition of Organic Compounds

- Fractionation during biosynthesis (lipids)

DeNiro and Epstein, 1977

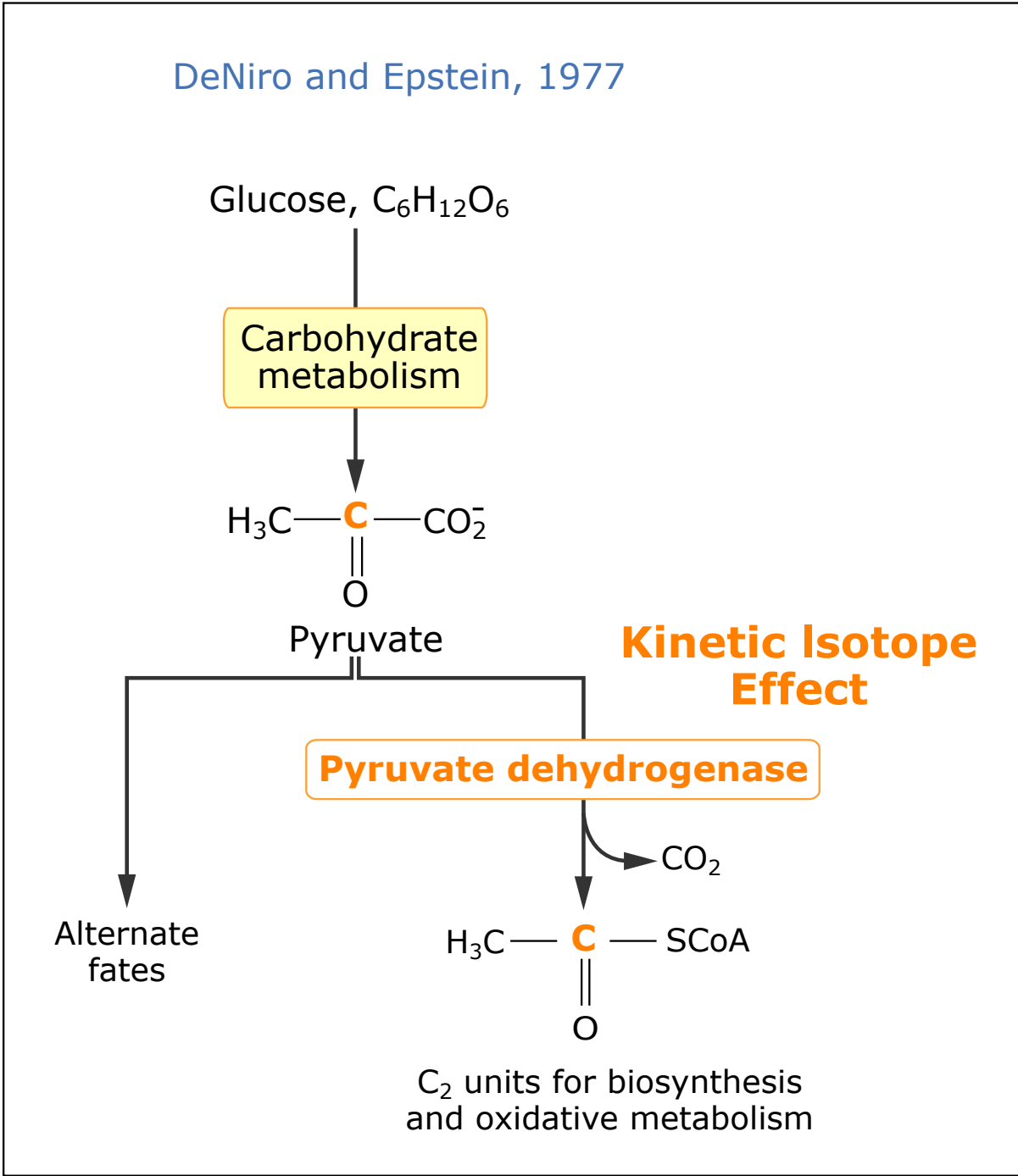
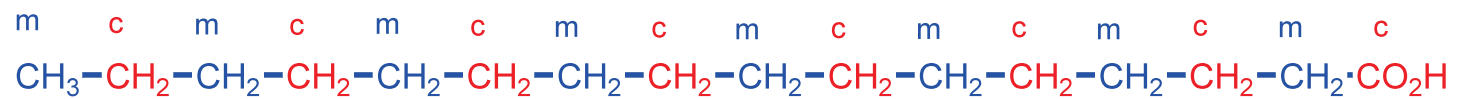


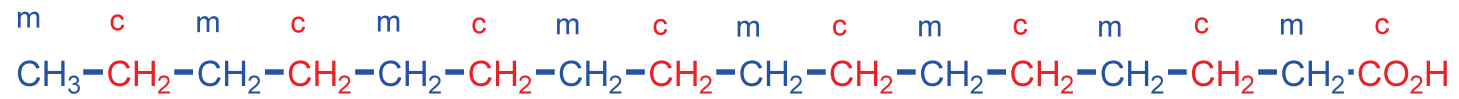
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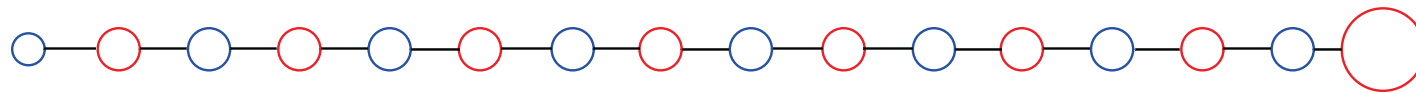
Product = C₁₆ fatty acid = 8 acetates



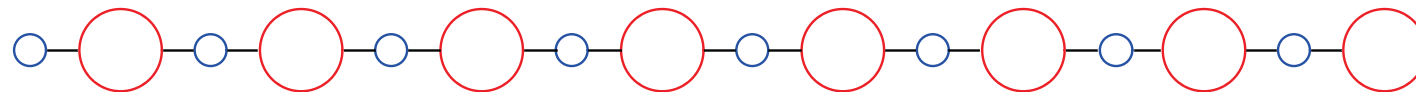
Methyl (m) and carboxyl (c) positions in acetate are converted to structurally equivalent CH₂ positions during biosynthesis



Isotopic equilibrium:



Equilibration within acetate → $\overset{m}{\text{blue}} - \overset{c}{\text{red}}$ Biosynthesis without subsequent equilibration →



Kinetic control throughout, acetate = $\overset{m}{\text{blue}} - \overset{c}{\text{red}}$

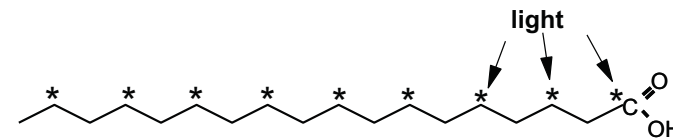
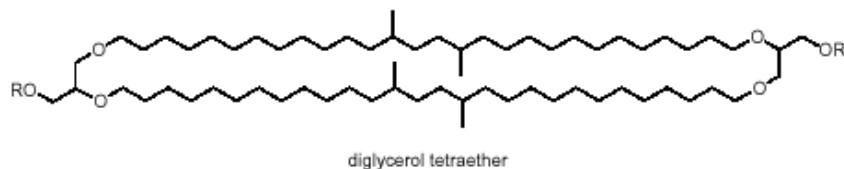
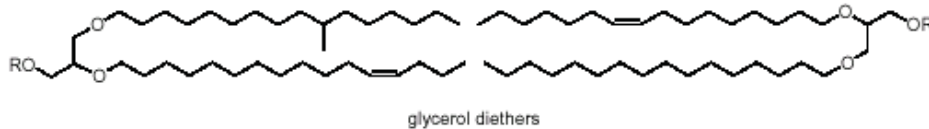
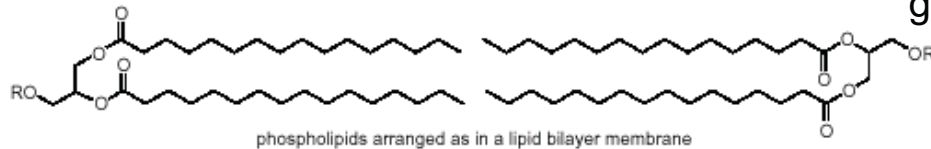
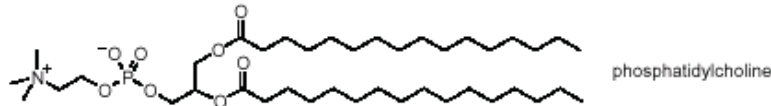
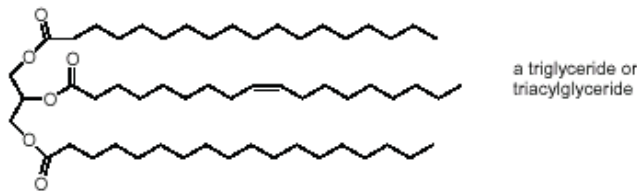
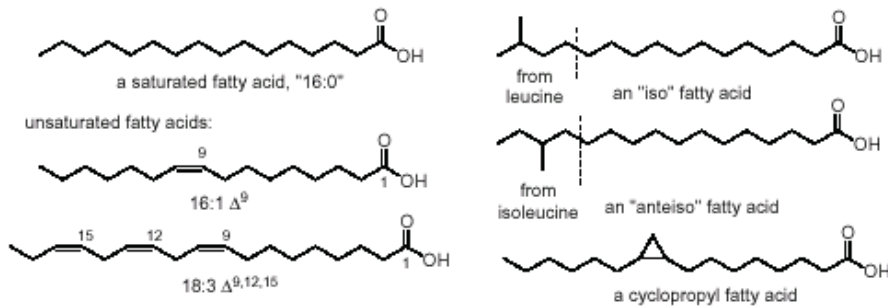


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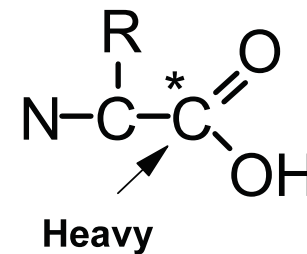
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Flows of C at the pyruvate branch point in the metabolism of *E. coli* grown aerobically on glucose (Roberts 1955). 74% of the pyruvate is decarboxylated to yield Ac-CoA. The observed depletion at odd-numbered positions of FAcids is shown at the right indicating that the isotope effect at C-2 in the pyruvate dehydrogenase reaction is 23‰

An important consequence of the pyruvate to acetate isotopic fractionation



Alternate carbons derived from acetate carboxyl down acetogenic lipidbackbones are light. In general lipids are also light, but not as light.



In contrast, the carboxyl carbon of amino acids is generally "heavy"

Two Origins for Isoprenoids

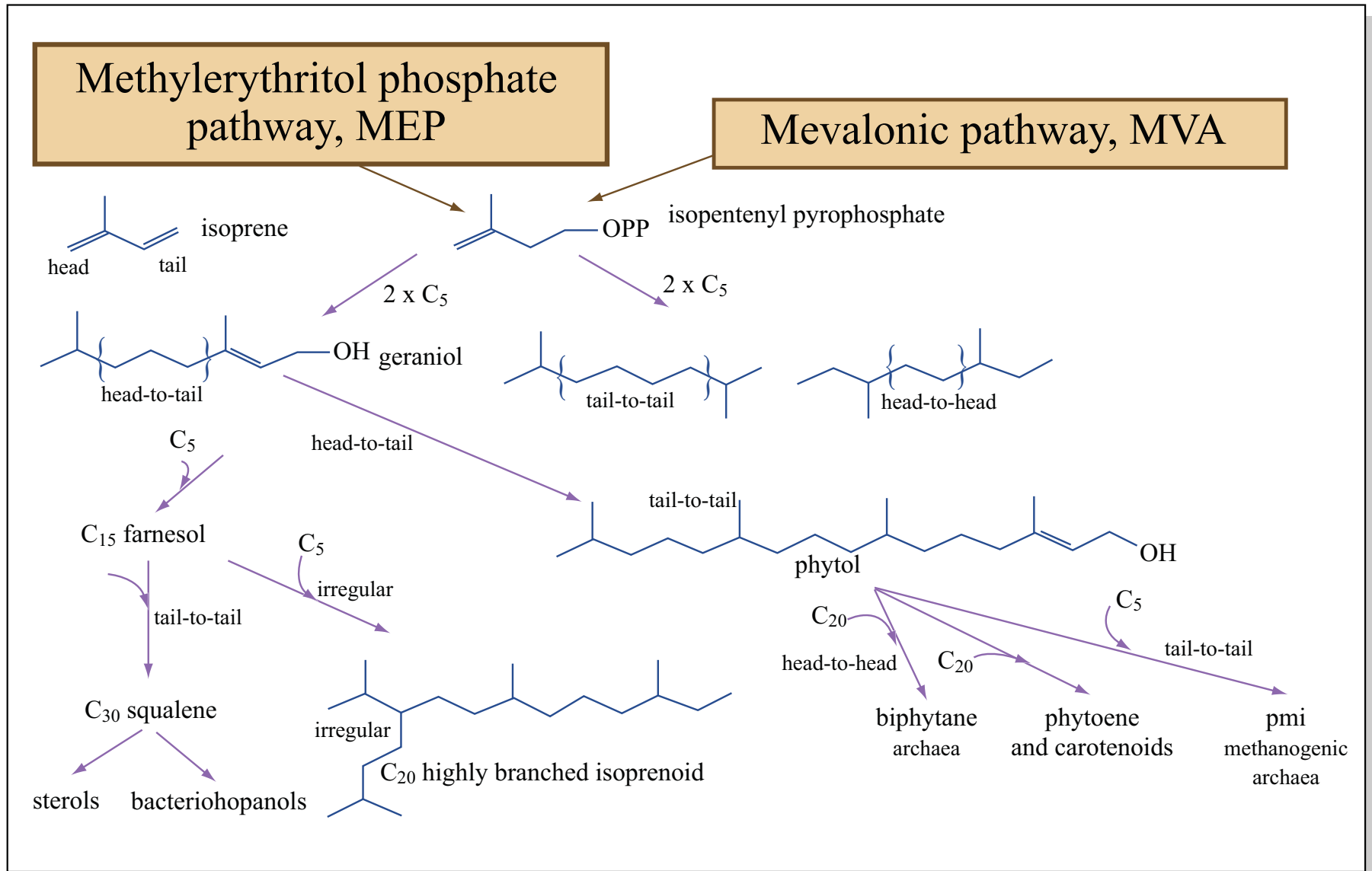
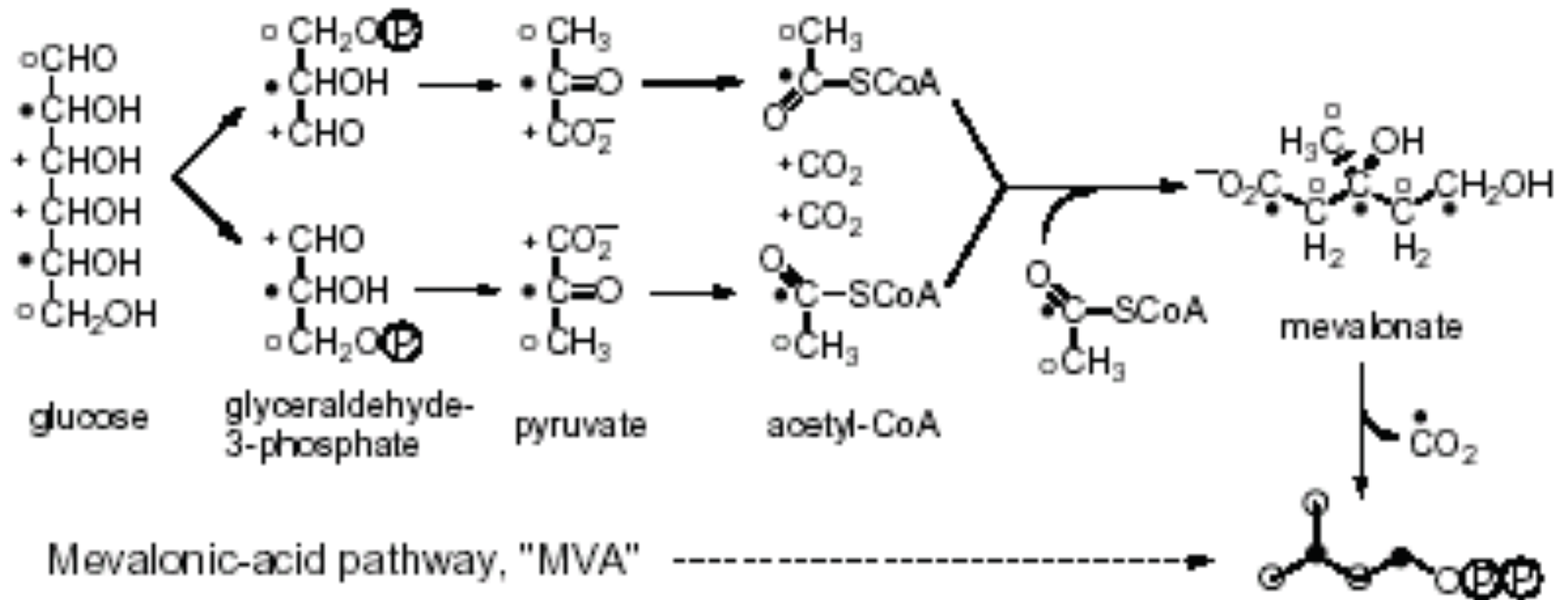
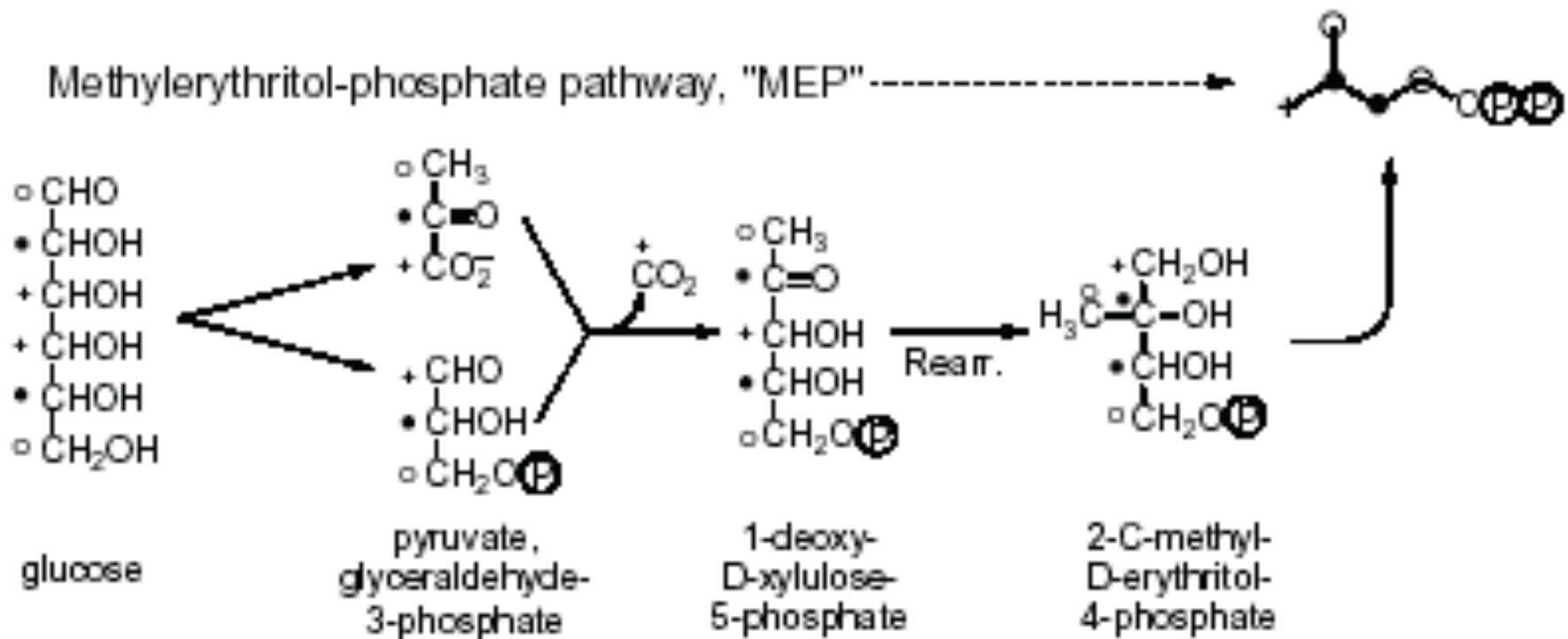


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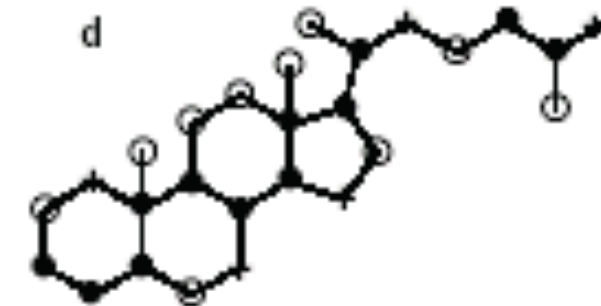
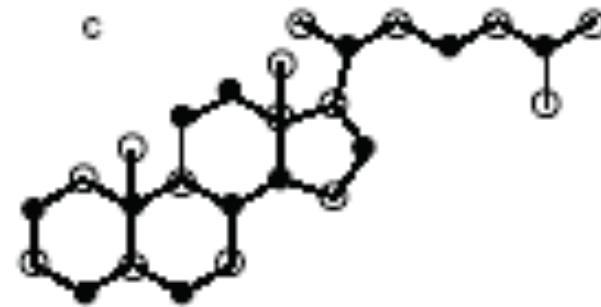
Labelling of isopentenylpyrophosphate from MVA pathway



Labelling of isopentenylpyrophosphate from MEP pathway



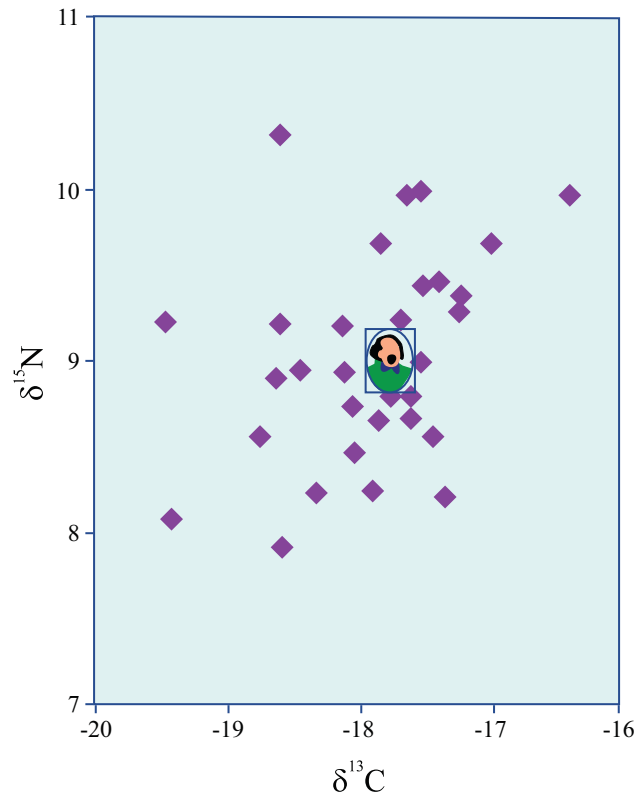
Labeling of phytol and cholesterol from MVA (a & c) and MEP (b & d)



Pathways Used for The Biosynthesis of Isoprenoid Lipids.

Organism	Pathway		Reference
Prokaryotes	-		Lange <i>et al.</i> , 2000
Bacteria	-		Boucher and Doolittle, 2000
Aquificales, Thermotogales	MEP		-
Photosynthetic bacteria	-		-
Chloroflexus	MVA		Rieder <i>et al.</i> , 1998
Chlorobium	MEP		Boucher and Doolittle, 2000
Gram positive eubacteria	-		-
Commonly	MEP		-
Streptococcus, Staphylococcus	MVA		Boucher and Doolittle, 2000
Streptomyces	MEP & MVA		Seto <i>et al.</i> , 1996
Spirochaetes	-		Boucher and Doolittle, 2000
Borrelia burgdorferi	MVA		-
Treponema pallidum	MEP		-
Proteobacteria	-		-
Commonly	MEP		-
Myxococcus, Nannocystis	MVA		Kohl <i>et al.</i> , 1983
Cyanobacteria	MEP		Disch <i>et al.</i> , 1998
Archaea	MVA		Lange <i>et al.</i> , 2000
Eukaryotes	-		-
Non-plastid-bearing	MVA		Lange <i>et al.</i> , 2000
Plastid-bearing	Plastid	Cytosol	-
Chlorophyta	MEP	MEP	Schwender <i>et al.</i> , 2001
Streptophyta	MEP	MVA	Lichtenthaler <i>et al.</i> , 1997
Euglenoids	MVA	MVA	Lichtenthaler 1999

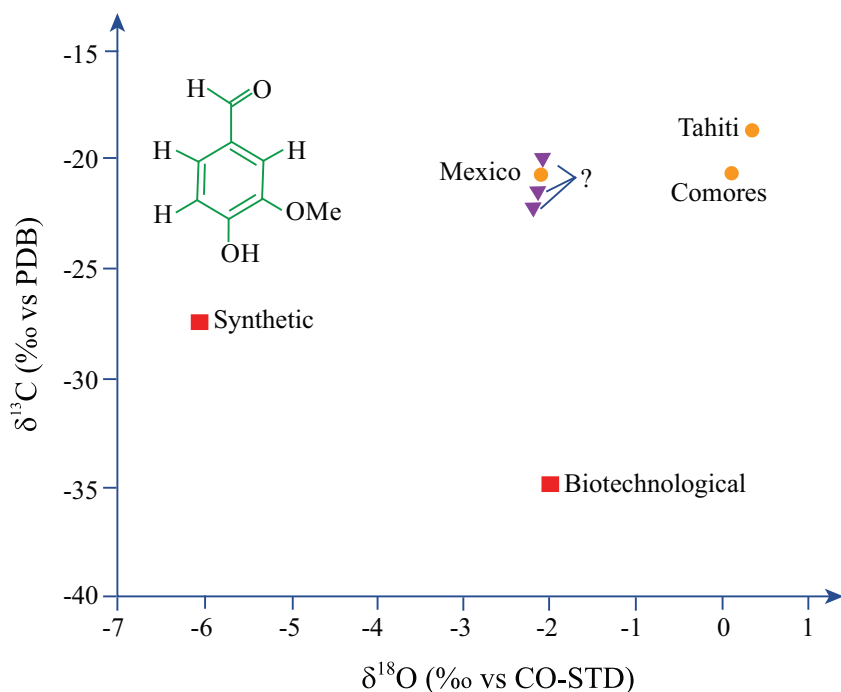
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Twin element stable isotope distributions ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$, in ‰ versus PDB and air, respectively) of hair samples taken from individual students at the University of Virginia (modified after Macko *et al.* [1998]). Hair is largely composed of the fibrous protein α -keratin. Its isotope composition is reflective of the recent diet of an individual, generally increasing slightly (1‰ in $\delta^{13}\text{C}$ and 3‰ in $\delta^{15}\text{N}$) with tropic level. The marked variability accords with the great diversity of modern diets. For comparison, George Washington's hair sample testifies to a rather balanced diet.

Figure by MIT OpenCourseWare.

Multi-element, Compound-specific Isotopic Analyses **Vanillin**



Dual isotopic ($\delta^{13}\text{C}$ and $\delta^{18}\text{O}$, in ‰ versus PDB and of standard CO, respectively) for the flavor compound vanillin. The three vanillin extracts from the naturally grown vanilla beans have similar $\delta^{13}\text{C}$ values, even though they come from geographically widely spaced sites: Mexico and the islands of the Comores and Tahiti. The Mexican sample, however, does differ markedly in $\delta^{18}\text{O}$, no doubt owing to major differences in $\delta^{18}\text{O}$ in the ambient water supply. Not surprisingly, major differences in both $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ are apparent in the synthetic and biotechnological products [Hener *et al.*, 1998]. On the basis of their dual isotopic values, the three samples of unknown origin can be assigned to Mexico.

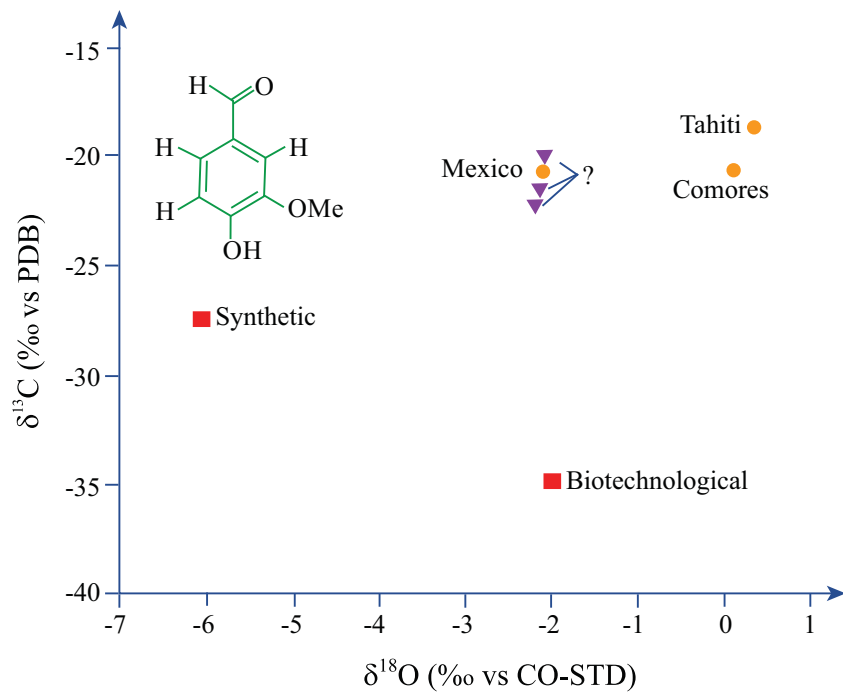
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Characterising Cocaine Sources

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Figure 2. Identification of geographic regions in South America where coca is commonly grown, based on dual isotope information of cocaine base as well as abundance of minor alkaloid components. Plotted on both axes are mixed expressions, each consisting of an isotope term and a concentration term; the y axis is $[\delta^{15}\text{N} \text{ cocaine } (\text{‰ versus air}) + 0.1 \times \text{relative concentration of truxilline } (\text{‰})]$, and the x axis is $[\delta^{13}\text{C} \text{ cocaine } (\text{‰ versus PDB}) - 10 \times \text{concentration of trimethoxycocaine}]$. Truxilline and trimethoxycocaine occur as two trace alkaloids in coca leaves. In addition to the obvious benefits for forensics, this illustration demonstrates the potential value of multi-isotope biomarker approaches for the geosciences to distinguish different geographical, climatological, and ecological regimes. Furthermore, it illustrates the importance of innovative data manipulation in biomarker research, especially when multiple isotope dimensions are employed. After *Ehleringer et al.* [2000].

Multi-element, Compound-specific Isotopic Analyses **Vanillin**



Dual isotopic ($\delta^{13}\text{C}$ and $\delta^{18}\text{O}$, in ‰ versus PDB and of standard CO, respectively) for the flavor compound vanillin. The three vanillin extracts from the naturally grown vanilla beans have similar $\delta^{13}\text{C}$ values, even though they come from geographically widely spaced sites: Mexico and the islands of the Comores and Tahiti. The Mexican sample, however, does differ markedly in $\delta^{18}\text{O}$, no doubt owing to major differences in $\delta^{18}\text{O}$ in the ambient water supply. Not surprisingly, major differences in both $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ are apparent in the synthetic and biotechnological products [Hener *et al.*, 1998]. On the basis of their dual isotopic values, the three samples of unknown origin can be assigned to Mexico.

Figure by MIT OpenCourseWare.

Reservoirs of carbon in the Atmosphere, Hydrosphere and Geosphere.

Reservoir	Reduced C	Mass, x 10 ¹⁸ moles Oxidized C	Total C
Atmosphere	-	0.06 ^a	0.06 ^a
Biosphere: plants and algae	0.13 ^b	-	0.13
Hydrosphere	-	3.3 ^c	3.3
Pelagic sediments	60 ^d	1300 ^d	1360
Continental margin sediments	>370 ^e	>1000 ^e	>1370
Sedimentary rocks	750 ^f	3500 ^f	4250
Crustal metamorphic and igneous rocks	100 ^g	?	>100
Mantle	-	-	27000 ^h

a- Holland (1984). b- Mopper and Degens (1979); Olson et al. (1985). c- Holland (1984).
d- Holser, et al. (1988). e- Minimum inventories required for C isotopic mass balance.
f- Ronov (1980). g- Hunt (1972). h- Derived from estimates of mantle mass and C concentration.

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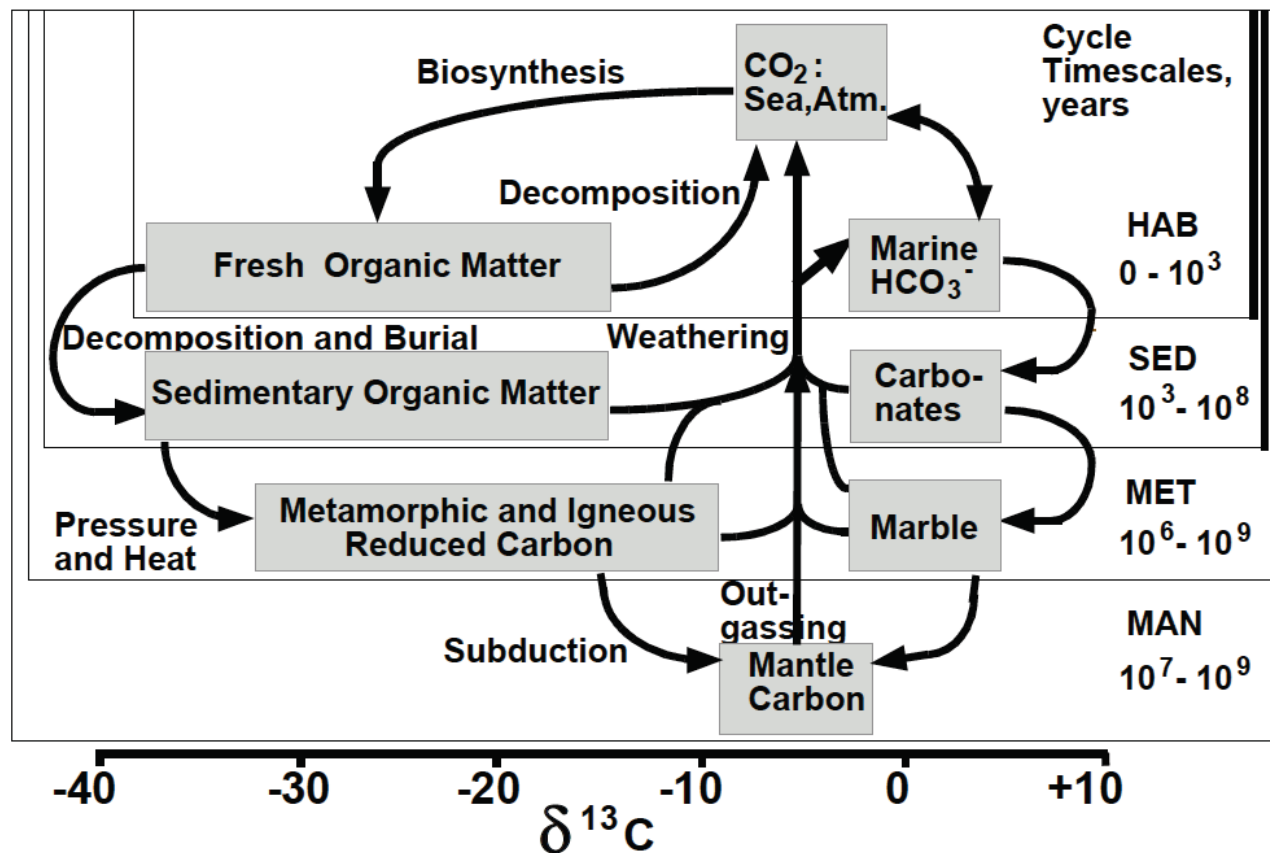


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Figure 1. Biogeochemical C cycle, showing principal C reservoirs (boxes) in the mantle, crust, oceans and atmosphere, and showing the processes (arrows) that unite these reservoirs. The range of each of these reservoir boxes along the horizontal axis gives a visual estimate of $\delta^{13}\text{C}$ values most typical of each reservoir. The vertical bars at right indicate the timeframes within which C typically completely traverses each of the four C sub-cycles (the HAB, SED, MET and MAN sub-cycles, see text). For example, C can traverse the hydrosphere-atmosphere-biosphere (HAB) sub-cycle typically in the time scale between 0 to 1000 years.

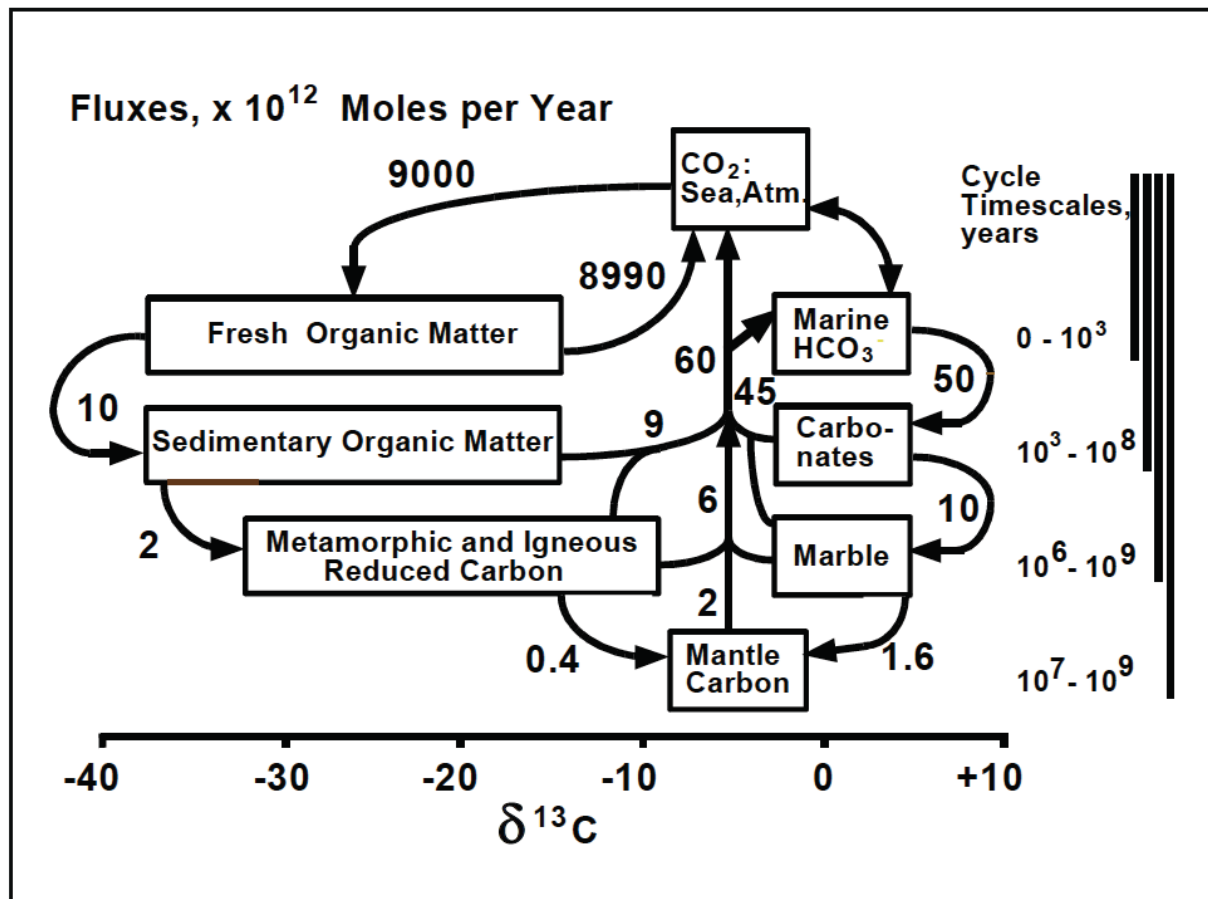


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Figure 2. Biogeochemical C cycle (as in Fig. 1), showing principal C reservoirs (boxes) and their isotopic compositions in the mantle, crust, oceans and atmosphere, and the processes (arrows) that unite these reservoirs. Numbers adjacent to the arrows give estimates of present-day fluxes, expressed in the units 10¹² mol yr⁻¹.

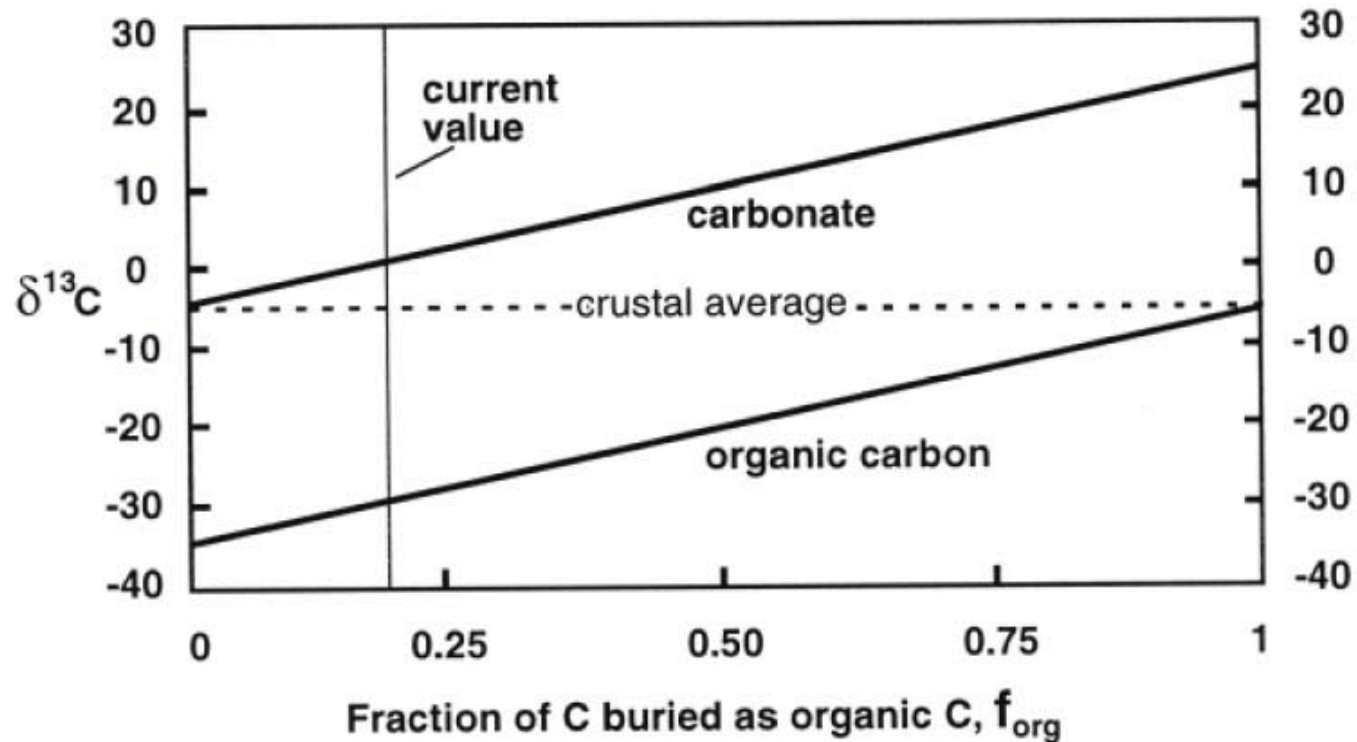


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Figure 3. Relationship between isotopic composition (δ_{carb} and δ_{org}) and the fraction of carbon buried as organic matter. The vertical separation between the lines depicts $\epsilon\Delta$, and thus reflects **the combined** effects of equilibria between inorganic C species and biological isotope discrimination (see text). A value $\epsilon_{\text{TOC}} = 30$ is depicted here, and represents the long-term average value during the past 800 Ma (Hayes et al. 1999). The vertical line represents the value of $f_{\text{org}} = 0.2$, which represents the current value for the global C cycle.

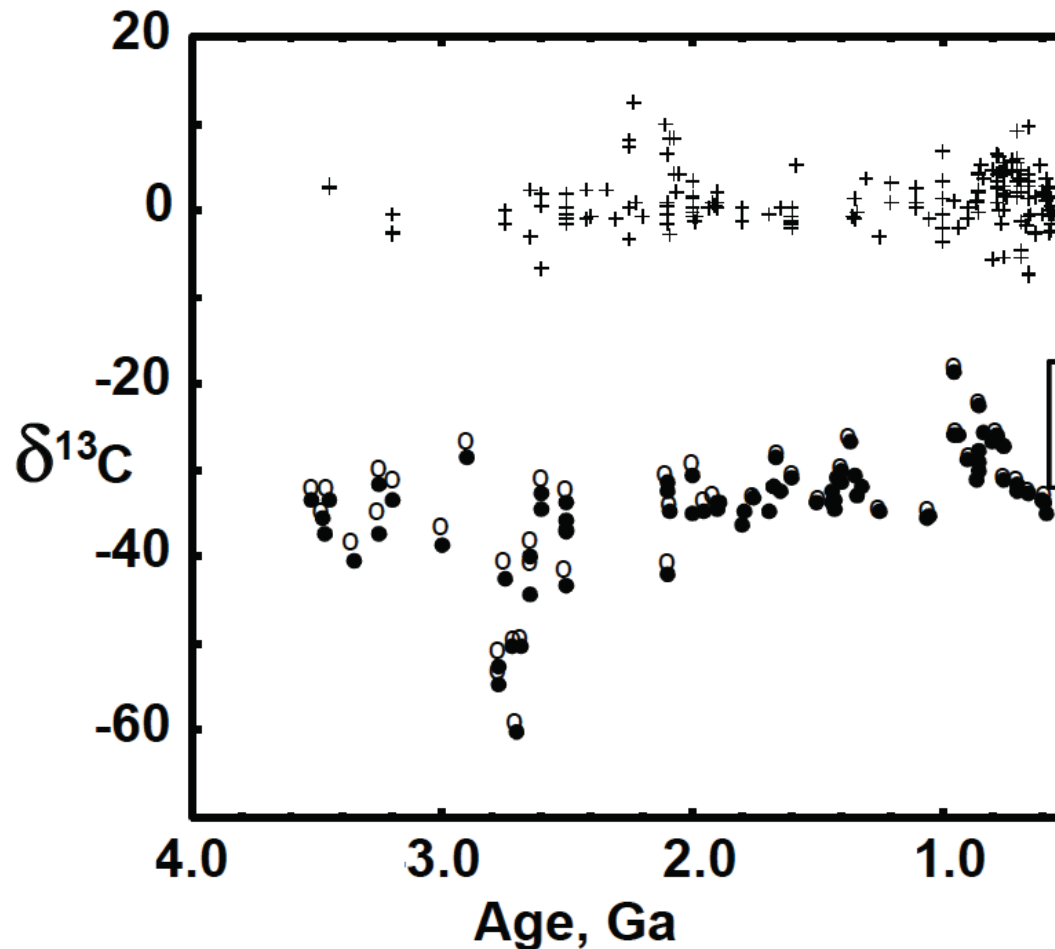
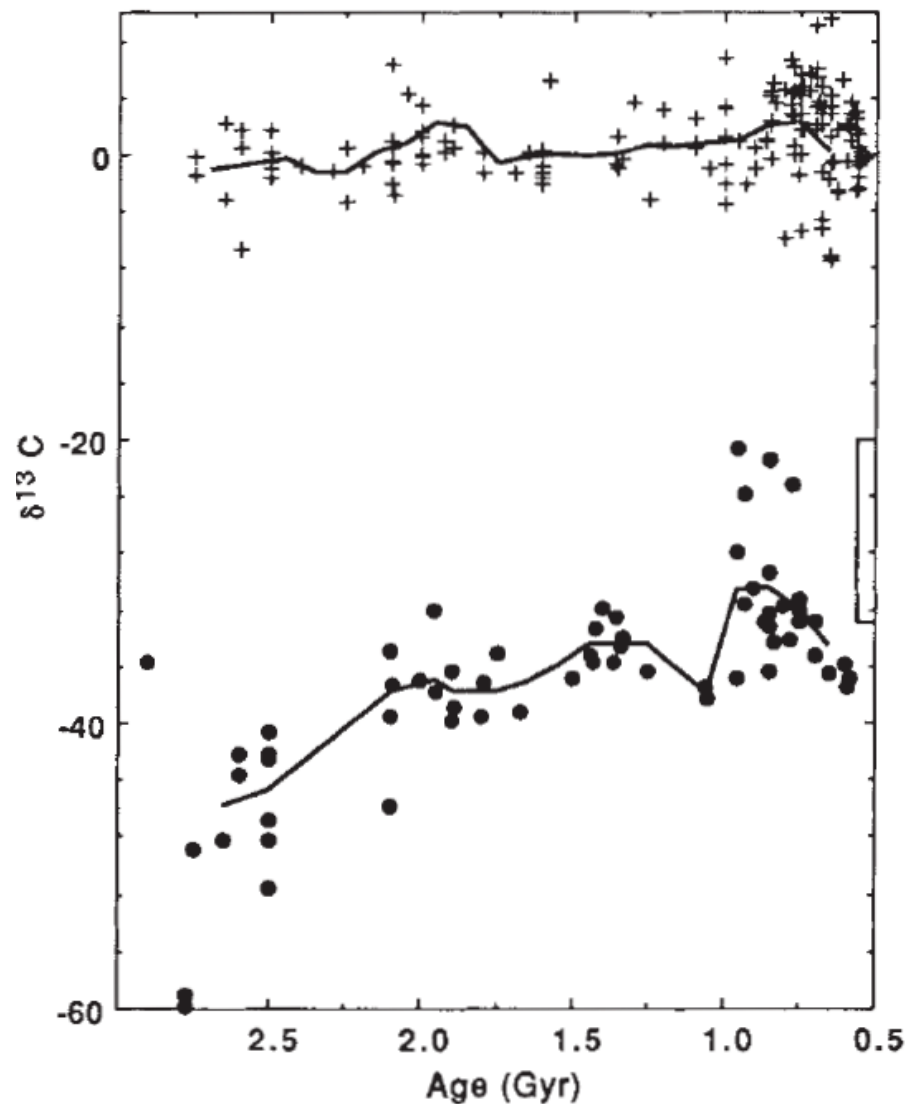


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Figure 5. Plot of age versus **δ_{carb} (crosses)** and **δ_{org}** for Archean and Proterozoic kerogens. Kerogen data (filled circles) are corrected for the effects of thermal alteration (Des Marais 1997a). Uncorrected data are shown as open circles. Between 2.2 to 2.0 billion years ago, note the high **δ_{carb} values** and the **virtual disappearance thereafter of δ_{org} values** more negative than -36. Other evidence indicates that atmospheric O₂ increased substantially at this time (see text).

Secular variation in $\delta^{13}\text{C}$ of purified kerogens (sedimentary organic matter)

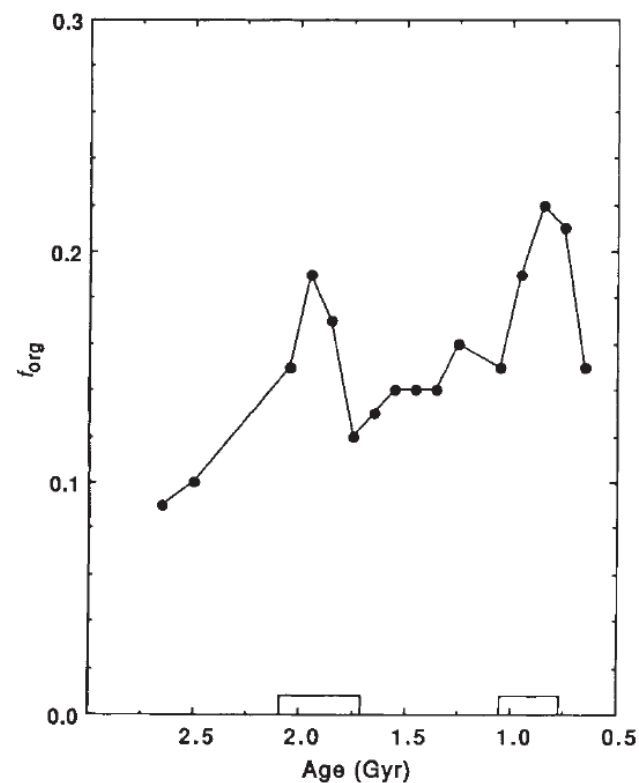


Carbon isotope evidence for the stepwise oxidation of the Proterozoic environment

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The oxidation of the Earth's crust and the increase in atmospheric oxygen early in Earth history have been linked to the accumulation of reduced carbon in sedimentary rocks. Trends in the carbon isotope composition of sedimentary organic carbon and carbonate show that during the Proterozoic aeon (2.5–0.54 Gyr ago) the organic carbon reservoir grew in size, relative to the carbonate reservoir. This increase, and the concomitant release of oxidizing power in the environment, occurred mostly during episodes of global rifting and orogeny.



Fraction of carbon buried as organic matter

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Source: David J. Des Marais, Harald Strauss, Roger E. Summons, J.M. Hayes. "Carbon Isotope Evidence for the Stepwise Oxidation of the Proterozoic environment." *Nature* 359 (1992): 605–9. doi:10.1038/359605a0.

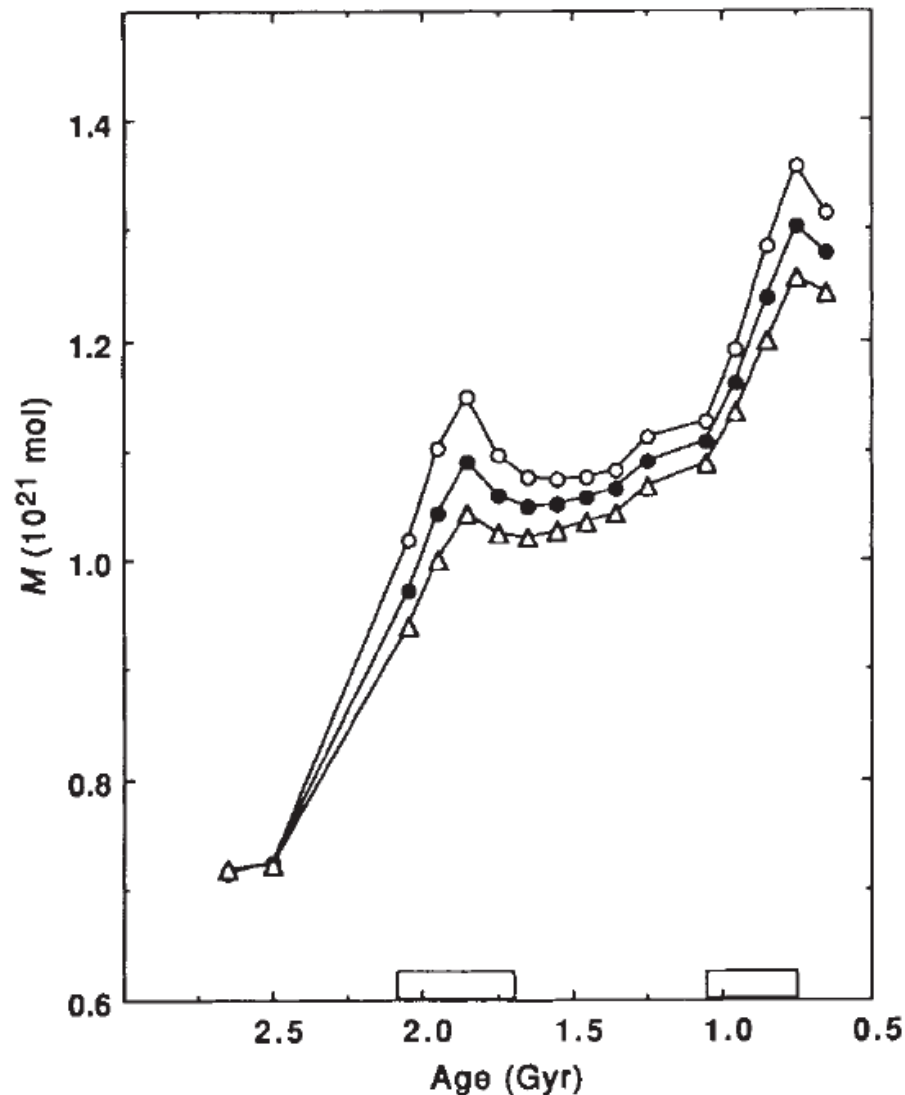


FIG. 4 Quantity of organic carbon in the crust (M) against age. Values of M are calculated according to equation (4) (in the text). Symbols represent calculations of M , assuming sediment half lives (see derivation of equation (4)) as follows : \circ , 300 Myr; \bullet , 400 Myr; \triangle , 500 Myr. Rectangles along the bottom margin depict time intervals of enhanced global rifting and orogeny (see text).

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ARTICLES

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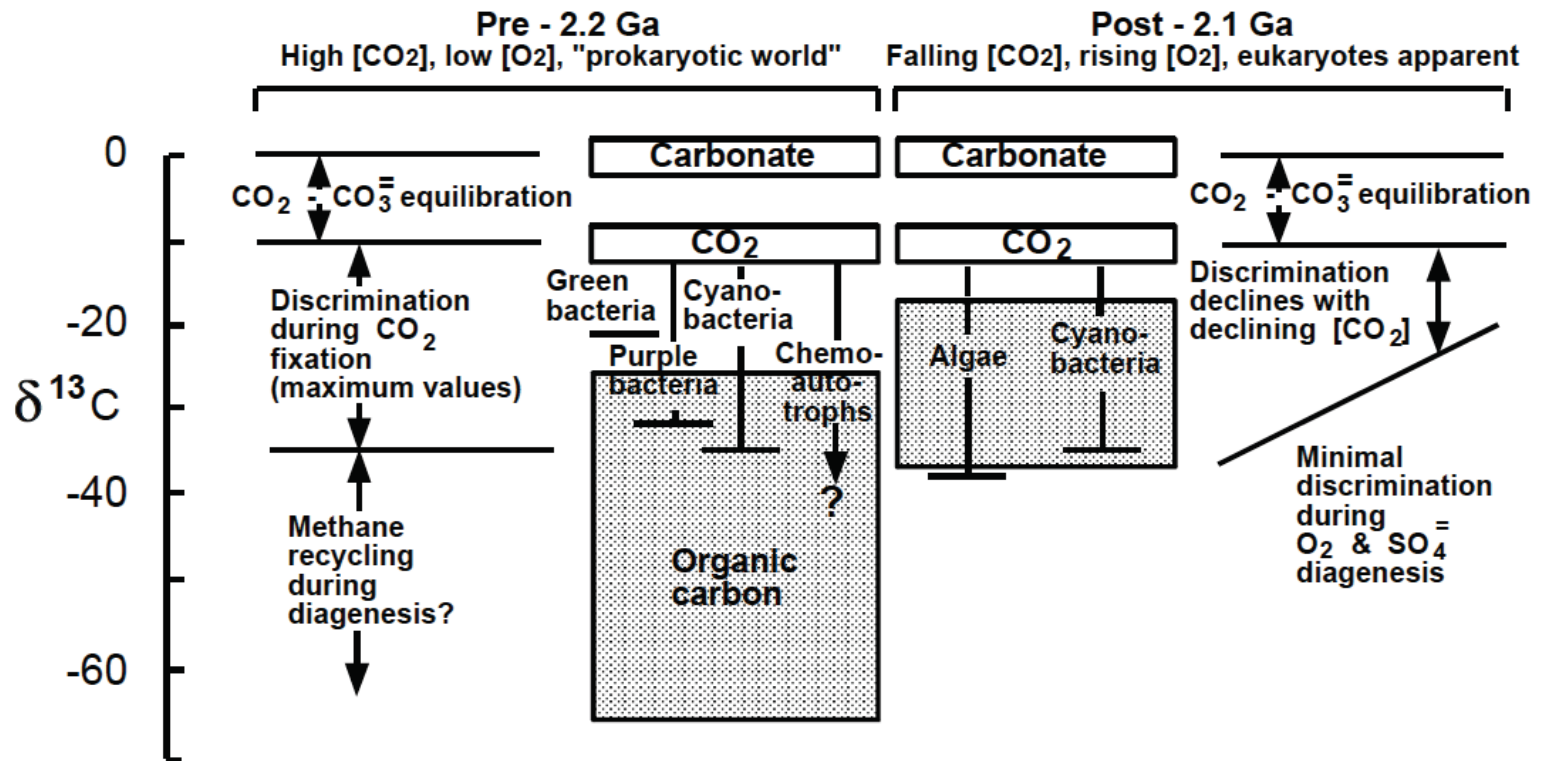


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Figure 8. Range of δ_{carb} and δ_{CO_2} values (open boxes) and δ_{org} values (shaded boxes), together with the processes proposed to explain their distribution prior to 2.2 Ga and subsequent to 2.1 Ga. A temperature of 15°C was assumed for the isotopic equilibrium between δ_{carb} and δ_{CO_2} . The lines associated with the various groups of autotrophic bacteria and algae illustrate the maximum discrimination expected for each group. The sloped line at right depicts declining discrimination over time, perhaps in response to declining CO₂ levels.

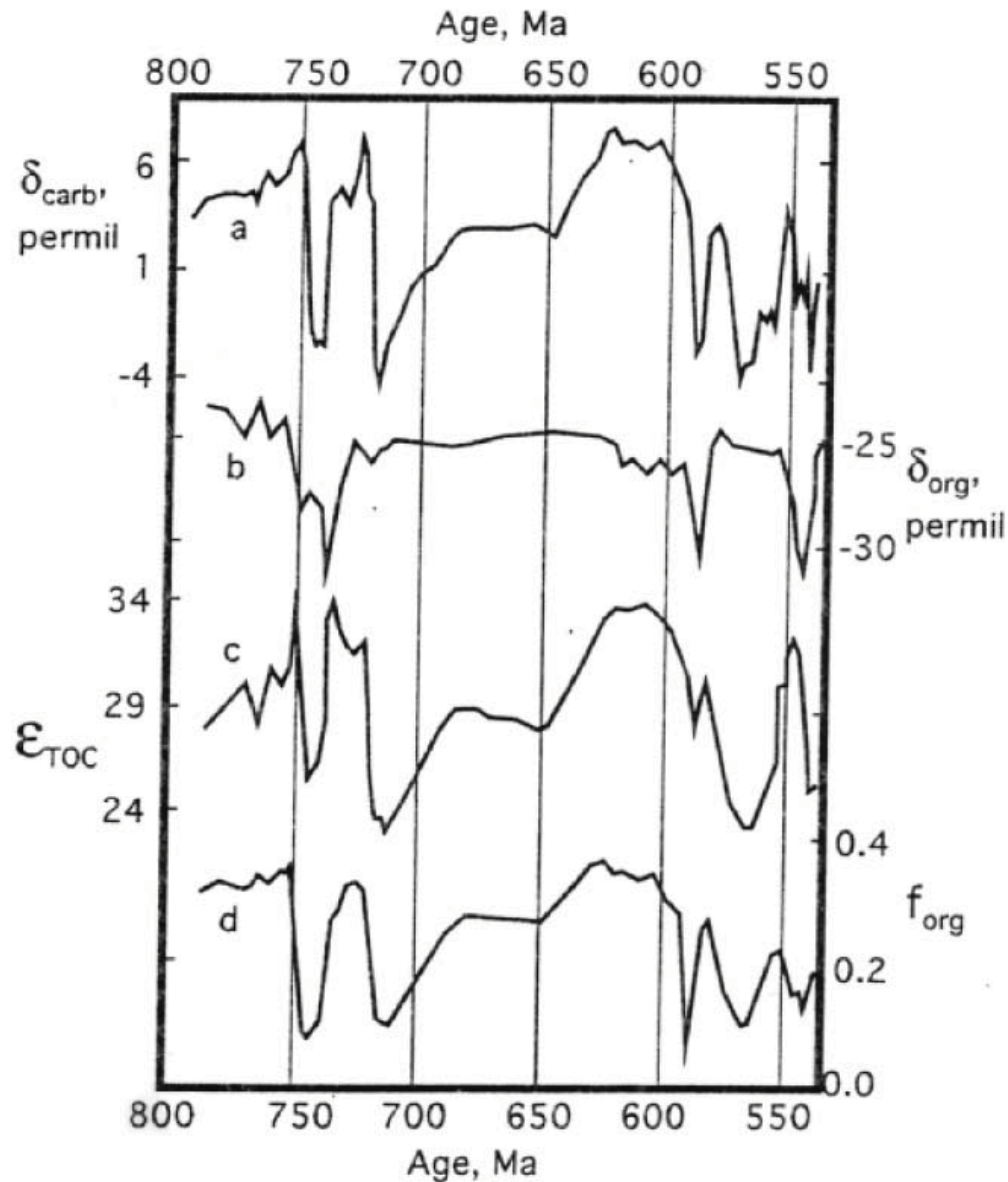


Figure 11. Neoproterozoic records of δ_{carb} (curve a) and δ_{org} (curve b) values (Hayes et al. 1999). Corresponding values for isotopic fractionation, ϵ_{TOC} and f_{org} during Neoproterozoic time are given by curves c and d, respectively. The periodic negative excursions are typically associated with glacial intervals (see text). Figure modified from Hayes et al. (1999).

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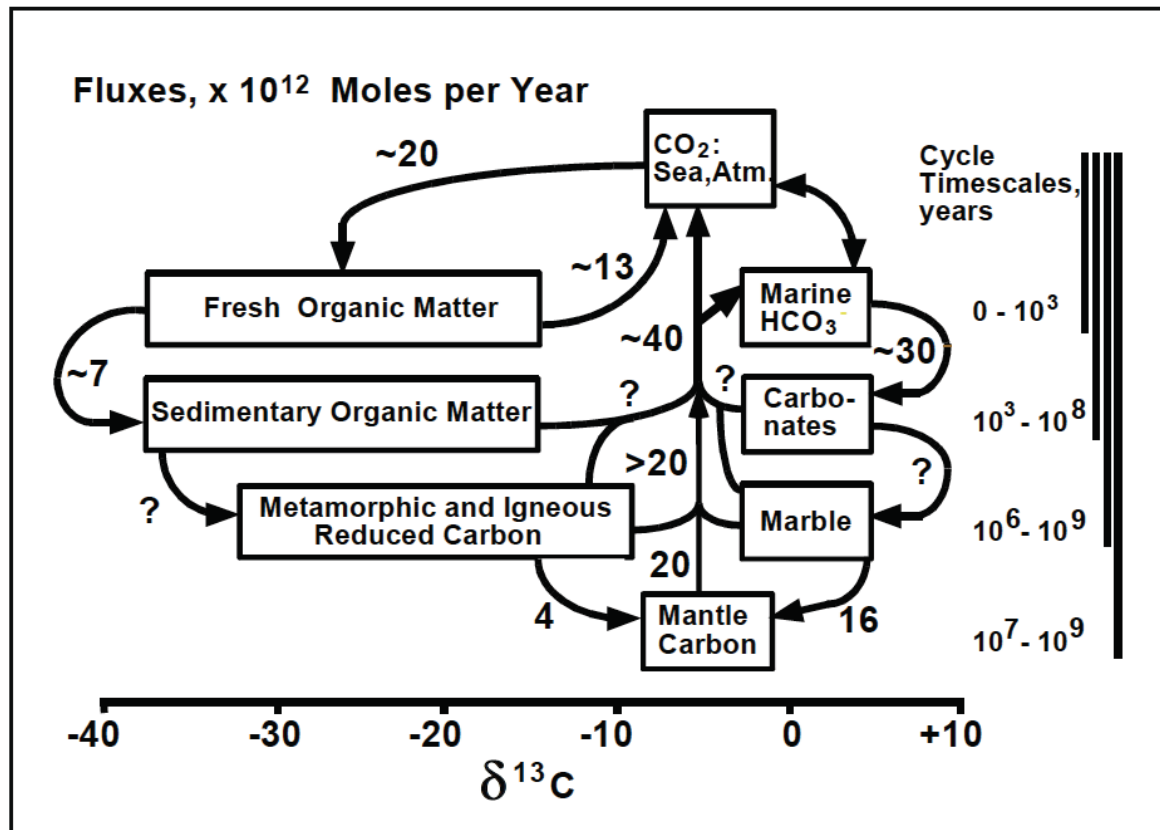
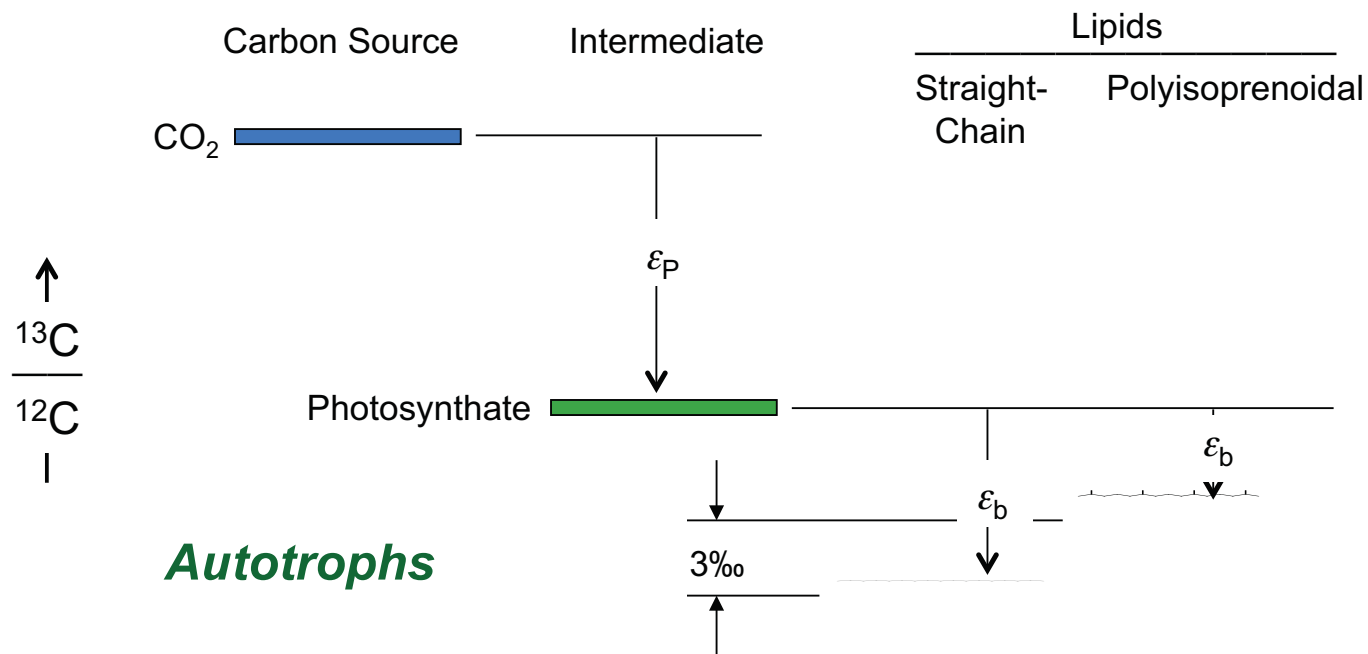
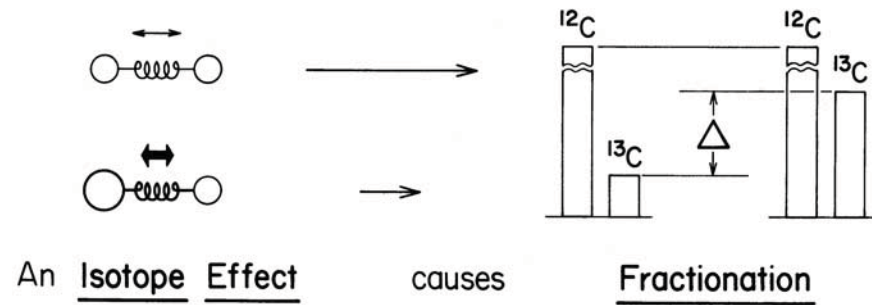


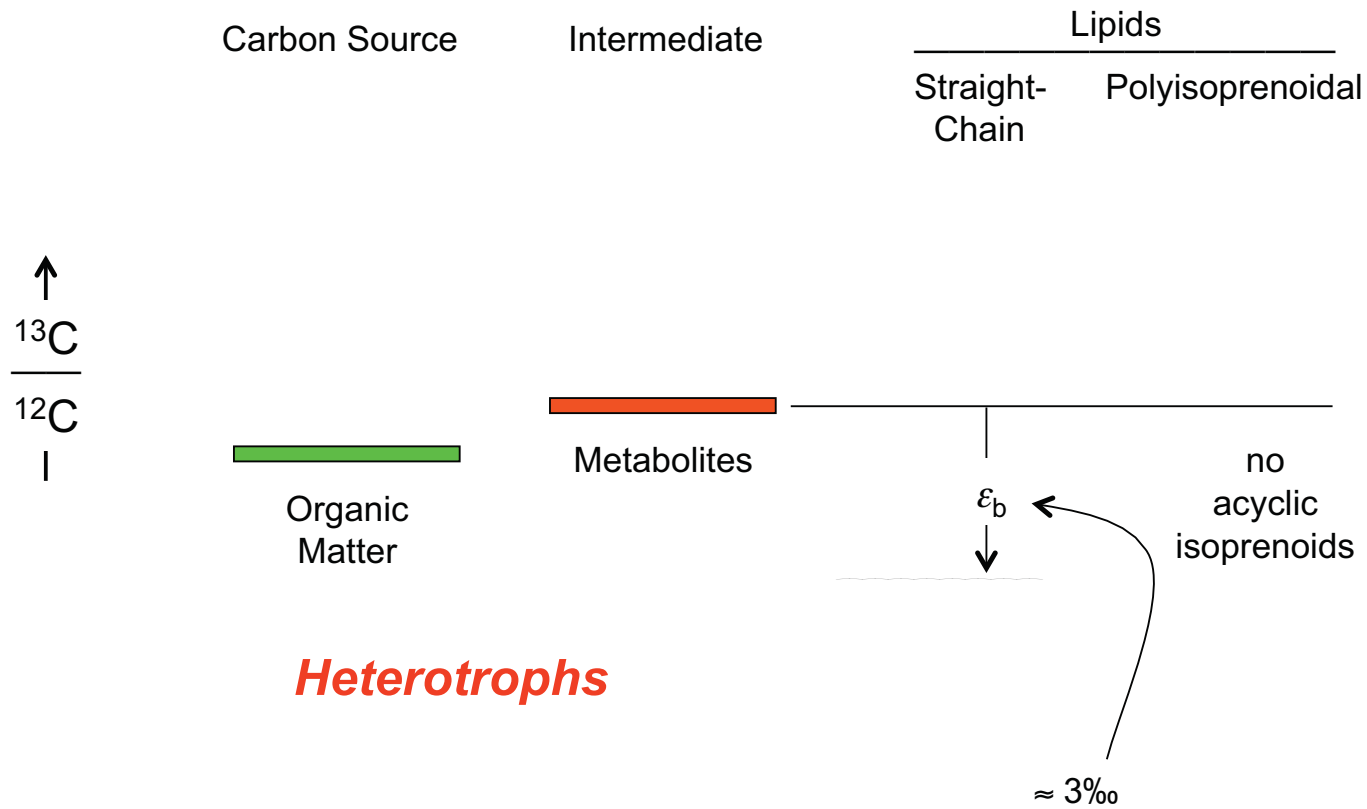
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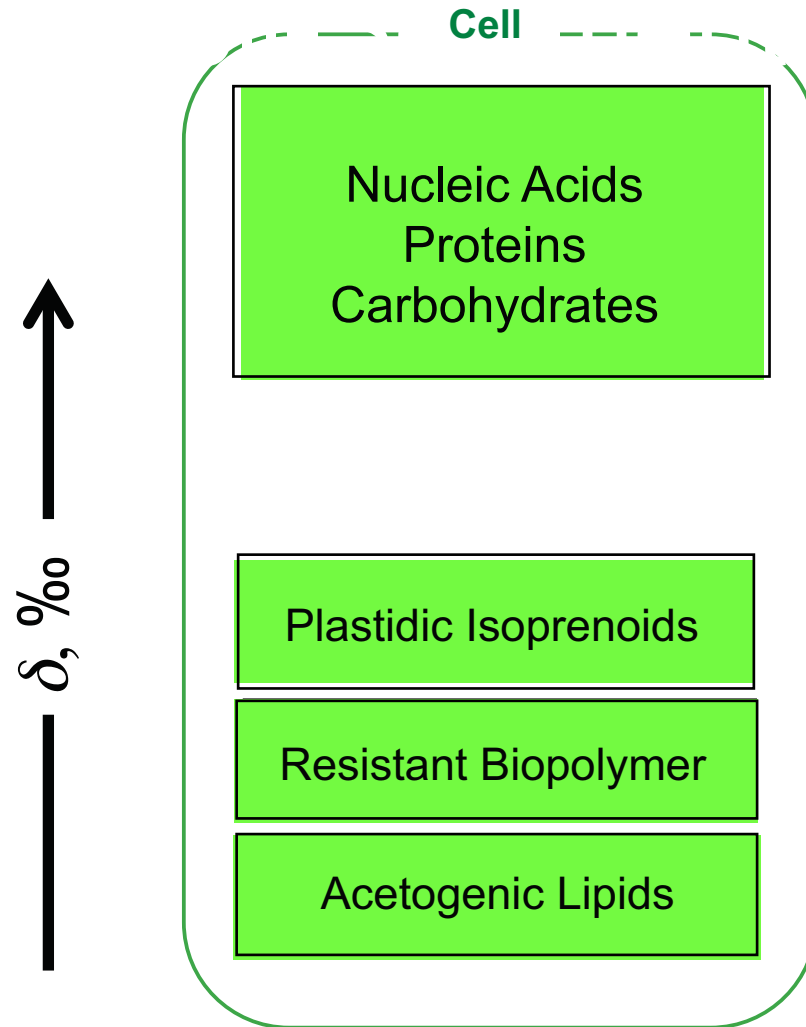
Figure 7. The biogeochemical C cycle prior to the advent of oxygenic photosynthesis, showing the much lower global primary productivity and the higher rates of thermal emanation of C (see text). Comparison with Figure 5 illustrates the enhancement of global primary productivity due to the development of oxygenic photosynthesis. Flux estimates are highly approximate, and are shown principally to illustrate the direction and magnitude of change over geologic time.

Biosynthesis of Organic Compounds



Biosynthesis of Organic Compounds





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